



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : C12N 15/12, C07K 14/755, A61K 38/37		A1	(11) International Publication Number: WO 97/03195 (43) International Publication Date: 30 January 1997 (30.01.97)
<p>(21) International Application Number: PCT/US96/11444</p> <p>(22) International Filing Date: 9 July 1996 (09.07.96)</p> <p>(30) Priority Data: 60/001,025 11 July 1995 (11.07.95) US</p> <p>(71) Applicant (for all designated States except US): CHIRON CORPORATION [US/US]; 4560 Horton Street, Emeryville, CA 94608 (US).</p> <p>(72) Inventors; and</p> <p>(75) Inventors/Applicants (for US only): HUNG, David, T. [US/US]; 460 Yerba Buena Avenue, San Francisco, CA 94127 (US). COHEN, Fred, E. [US/US]; 767 Rhode Island, San Francisco, CA 94107 (US). INNIS, Michael [US/US]; 315 Constance Place, Moraga, CA 94556 (US).</p> <p>(74) Agents: CHUNG, Ling-Fong; Chiron Corporation, Intellectual Property - R440, P.O. Box 8097, Emeryville, CA 94662-8097 (US) et al.</p>		<p>(81) Designated States: AU, CA, CN, IL, JP, MX, US, European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE).</p> <p>Published <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i></p>	
<p>(54) Title: NOVEL FACTOR VIII:C POLYPEPTIDE ANALOGS WITH ALTERED PROTEASE SITES</p> <p>(57) Abstract</p> <p>Factor VIII:C polypeptide analogs are provided that are native Factor VIII:C polypeptides that contain amino acid modifications at one or more amino acid residues adjacent an Arg residue, provided that the Arg residue is not at a Factor VIII:C activation site. Such modifications create one or more Arg-Pro or Pro-Arg linkages. Nucleic acid molecules encoding Factor VIII:C polypeptide analogs, vectors and host cells containing such nucleic acid molecules are also provided. Additional modifications include the creation of a tripeptide having the formula P₃-P₂-P₁, wherein P₃ is a residue selected from the group consisting of Phe, Glu, and Pro; P₂ is any amino acid residue except Ser and P₂ is not Leu335 and is not Asn1720; and P₁ is Arg. Other modifications include substitutions at the non-activating Arg residues occurring at positions Arg336, Arg1719 and/or Arg1721. Further provided are analog complexes that contain at least two such analogs. Methods of producing the analogs, the analog complexes, nucleic acids encoding the same, vectors, and host cells are also provided as well as methods of using such compositions for prevention or treatment of active Factor VIII:C polypeptide deficiencies.</p>			

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NOVEL FACTOR VIII:C POLYPEPTIDE ANALOGS WITH
ALTERED PROTEASE SITES

Field of the Invention

5 This invention relates to the discovery that an active Factor VIII:C polypeptide analog can be made that is modified at a site adjacent to an arginine ("Arg") residue, where the Arg is at a site other than at an activating site, creating an arginine-proline ("Arg-Pro")
10 linkage or a proline-arginine ("Pro-Arg") linkage. This invention also relates to a Factor VIII:C polypeptide analog with a modification that comprises creation of a tripeptide having the formula $P_3-P_2-P_1$, wherein P_3 is a residue selected from the group consisting of Phe, Glu, and Pro; P_2 is any amino acid residue except Ser and P2 is not Leu335 and is not Asn1720; and P_1 is Arg.
15 Additionally, the invention pertains to substitutions at the non-activating Arg residues occurring at positions Arg336, Arg1719 and/or Arg1721. This invention further
20 relates to an analog complex comprising at least two such analogs, or one such analog and a native Factor VIII:C polypeptide, nucleic acid molecules encoding such analogs, vectors and host cells comprising the nucleic acid molecules, pharmaceutical compositions comprising
25 the analogs or analog complexes, methods of making the analogs, nucleic acid molecules, vectors and host cells, and methods of prevention or treatment of active Factor VIII:C deficiency using the analogs, complexes, and/or nucleic acid molecules, vectors and host cells.

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Background of the Invention

Hemophilia A is an X-chromosome-linked inherited bleeding diathesis that results from the

deficiency of an active blood clotting factor termed Factor VIII:C. The disease afflicts approximately 1 in 10,000 males. Factor VIII:C is a large glycoprotein that participates in the blood coagulation cascade that 5 ultimately converts soluble fibrinogen to insoluble fibrin clot, effecting hemostasis.

The deduced primary amino acid sequence of human Factor VIII:C determined from the cloned cDNA indicates that Factor VIII:C is a heterodimer processed 10 from a larger precursor polypeptide consisting of 2351 amino acids, referred to herein as the precursor or full-length Factor VIII:C molecule, of which the first 19 N-terminal residues comprise the signal sequence. Therefore, the mature Factor VIII:C molecule, starting 15 with Ala1, which does not contain the signal peptide sequence, includes a sequence of 2332 amino acids. Amino acids from about 1 to about 1648 of the mature Factor VIII:C molecule give rise to "heavy chain" fragments with molecular weights ranging from approximately 90 kD to 200 20 kD. Amino acids from about 1649 to about 2331 of the mature Factor VIII:C molecule comprise a "light chain" with a molecular weight of approximately 80 kD ("the 80 kD subunit"). The heterodimeric mature Factor VIII:C molecule consists of the heavy and light chains 25 associated by a metal ion bridge, and lacking amino acids 741-1648 (the B domain).

The mature Factor VIII:C molecule consists of a triplicated A domain of 330 amino acids, a unique B domain of 980 amino acids, and a duplicated C domain of 30 150 amino acids with the structure NH₂-A1-A2-B-A3-C1-C2-COOH. See, e.g., Kaufman, R. J., *Structure and Biology of Factor VIII*, in Part VI, *Hemostasis and Thrombosis*, pp. 1276-1284; and Pan et al., *Nature Structural Biology* (1995) 2:740-744.

35 Factor VIII:C is known to be activated by plasma proteases such as thrombin. During activation, the mature Factor VIII:C polypeptide is cleaved to

generate heavy and light chain fragments that are further cleaved. For example, cleavage of the light chain after arginine residue 1689 ("Arg1689") yields a light chain fragment of about 73 kD ("the 73 kD fragment"), and 5 cleavage of the heavy chain after arginine residue 372 ("Arg372") yields smaller heavy chain fragments of about 50 kD and 43 kD ("the 50 kD and 43 kD fragments," respectively), as described in Eaton et al. (1986), *Biochem.* 25: 505-512. At a minimum, the complex formed 10 by the 50 kD and 73 kD polypeptides appears to be required for Factor VIII:C coagulant activity. Following activation, the heavy and light chain fragments of Factor VIII:C are inactivated by plasma proteases.

Patients suffering from hemophilia A are 15 conventionally treated with purified or substantially purified Factor VIII:C. A difficulty in such treatment is the relatively short half-life of externally administered Factor VIII:C, lasting about 8 to 12 hours. This instability of Factor VIII:C derives in part from 20 its susceptibility to proteolytic cleavage by plasma protease. Such plasma proteases, for example, thrombin, Factor Xa, and activated protein C ("APC"), inactivate Factor VIII:C by cleaving the molecule at multiple sites.

It would be advantageous, therefore, to produce 25 Factor VIII:C polypeptides with improved properties.

Summary of the Invention

It is, therefore, an object of the present invention to provide a Factor VIII:C polypeptide analog 30 that has improved properties.

In accordance, therewith, there is provided an active Factor VIII:C polypeptide analog that is substantially the same as a native Factor VIII:C polypeptide, except for modification at a site that is 35 adjacent to a non-activating Arg residue, that is, adjacent to an Arg residue that is not at an activation site, such as Arg1689 or Arg372. The modification of the

present invention creates an Arg-Pro linkage or a Pro-Arg linkage. Such a modification can be achieved by one or more amino acid substitutions, additions or deletions.

In accordance with another object of the present invention, there is provided an analog as above where the non-activating Arg residue is at least one selected from the group consisting of amino acid residues 220, 226, 250, 279, 282, 336, 359, 562, 747, 776, 1310, 1313, 1645, 1648, 1719, and 1721, numbered with respect to the native Factor VIII:C polypeptide sequence, as depicted in Figures 1A-1F.

In accordance with another object of the present invention, there is provided an analog with a modification that comprises creation of a tripeptide having the formula $P_3-P_2-P_1$, wherein P_3 is a residue selected from the group consisting of Phe, Glu, and Pro; P_2 is any amino acid residue except Ser and P_2 is not Leu335 and is not Asn1720; and P_1 is Arg.

In accordance with another object of the invention, there is provided an active Factor VIII:C polypeptide analog comprising a native Factor VIII:C polypeptide that is modified at at least one non-activating Arg residue selected from the group consisting of Arg336, Arg1719 and Arg1721, wherein the modification comprises a substitution of any of amino acids Pro, Glu, Asp, Asn, Gln, Ser and Tyr for Arg336, a substitution of any of amino acids Pro, Glu, Asp, Asn, Gln, Ser and Tyr for Arg1719 and/or a substitution of any of amino acids Glu, Asp, Asn, Gln, Ser and Tyr for Arg1721, numbered with respect to the native Factor VIII:C polypeptide sequence.

In accordance with a further object of the present invention, there is provided an analog as above where the native Factor VIII:C polypeptide that is modified is selected from the group consisting of (a) a full-length Factor VIII:C molecule comprising a signal peptide and all A, B, and C domains; (b) a native Factor

VIII:C molecule comprising all A, B, and C domains and lacking a signal peptide; (c) a truncated Factor VIII:C molecule lacking a signal peptide and at least a portion of the B domain; (d) a cleaved Factor VIII:C molecule containing a light chain subunit of molecular weight of about 80 kD; (e) a cleaved Factor VIII:C molecule containing a heavy chain fragment of molecular weight in a range of about 90 kD to about 200 kD; (f) a cleaved Factor VIII:C molecule comprising a heavy chain fragment of a molecular weight of about 90 kD; (g) a cleaved Factor VIII:C molecule containing a heavy chain fragment of molecular weight of about 50 kD, (h) a cleaved Factor VIII:C molecule containing a heavy chain fragment of molecular weight of about 43 kD; and (i) a cleaved Factor VIII:C molecule containing a light chain fragment of molecular weight of about 73 kD.

In accordance with a further object of the present invention, there is provided an active Factor VIII:C analog complex that contains either at least two Factor VIII:C polypeptide analogs as above, or a Factor VIII:C polypeptide analog and a Factor VIII:C polypeptide, together with a metal ion. For example, the analog complex herein can comprise two Factor VIII:C polypeptide analogs, and the two analogs can be selected from the group consisting of analogs of molecular weights of about (a) 80 kD and 90 kD; (b) 73 kD and 90 kD; (c) 80 kD and 50 kD; (d) 80 kD and 43 kD; (e) 73 kD and 50 kD; and (f) 73 kD and 43 kD.

In accordance with yet another object of the present invention, there is provided a method of producing a Factor VIII:C polypeptide analog as above by (a) providing a native Factor VIII:C polypeptide that contains an amino acid sequence, and (b) modifying at least one amino acid residue in the amino acid sequence to produce an analog as above.

In accordance with still another object of the present invention, there is provided a nucleic acid

present invention creates an Arg-Pro linkage or a Pro-Arg linkage. Such a modification can be achieved by one or more amino acid substitutions, additions or deletions.

In accordance with another object of the present invention, there is provided an analog as above where the non-activating Arg residue is at least one selected from the group consisting of amino acid residues 220, 226, 250, 279, 282, 336, 359, 562, 747, 776, 1310, 1313, 1645, 1648, 1719, and 1721, numbered with respect to the native Factor VIII:C polypeptide sequence, as depicted in Figures 1A-1F.

In accordance with another object of the present invention, there is provided an analog with a modification that comprises creation of a tripeptide having the formula $P_3-P_2-P_1$, wherein P_3 is a residue selected from the group consisting of Phe, Glu, and Pro; P_2 is any amino acid residue except Ser and P_2 is not Leu335 and is not Asn1720; and P_1 is Arg.

In accordance with another object of the invention, there is provided an active Factor VIII:C polypeptide analog comprising a native Factor VIII:C polypeptide that is modified at at least one non-activating Arg residue selected from the group consisting of Arg336, Arg1719 and Arg1721, wherein the modification comprises a substitution of any of amino acids Pro, Glu, Asp, Asn, Gln, Ser and Tyr for Arg336, a substitution of any of amino acids Pro, Glu, Asp, Asn, Gln, Ser and Tyr for Arg1719 and/or a substitution of any of amino acids Glu, Asp, Asn, Gln, Ser and Tyr for Arg1721, numbered with respect to the native Factor VIII:C polypeptide sequence.

In accordance with a further object of the present invention, there is provided an analog as above where the native Factor VIII:C polypeptide that is modified is selected from the group consisting of (a) a full-length Factor VIII:C molecule comprising a signal peptide and all A, B, and C domains; (b) a native Factor

composition that contains an active Factor VIII:C polypeptide analog or analog complex as above, and a pharmaceutically acceptable excipient.

Furthermore, there is also provided, in accordance with another object of the present invention, methods for prevention or treatment of active Factor VIII:C deficiency in a mammal comprising administering thereto a therapeutically effective amount of (a) an active Factor VIII:C polypeptide analog as above, or (b) an active Factor VIII:C polypeptide analog complex as above, or (c) a nucleic acid molecule as above, or (d) a recombinant vector as above, or (e) a nucleic acid molecule as above together with an active Factor VIII:C polypeptide analog, or (f) a recombinant vector as above together with an active Factor VIII:C polypeptide analog.

Further objects, features, and advantages of the present invention will become apparent from the following detailed description. It should be understood, however, that the detailed description, while indicating the preferred embodiments of the invention, is given by way of illustration only, since various changes and modifications within the spirit and scope of the invention will become apparent to those skilled in the art from this detailed description.

Brief Description of the Figure

Figures 1A-1F depict the amino acid sequence of native Factor VIII:C.

30

Detailed Description of the Invention

The invention described herein draws on previously published work and pending patent applications. By way of example, such work consists of 35 scientific papers, patents or pending patent applications. All of these publications and

applications, cited previously or below are hereby incorporated by reference.

The inventors herein have discovered that Factor VIII:C polypeptide analogs can be made that have 5 improved properties. These analogs include one or more amino acid residues that are modified from the native structure. The modification can be at least one amino acid substitution, addition or deletion, at an amino acid residue adjacent to an Arg residue so as to generate, for 10 example, an Arg-Pro linkage or a Pro-Arg linkage.

Furthermore, the Factor VIII:C polypeptide analog can also include a modification that comprises creation of a tripeptide having the formula $P_3-P_2-P_1$, wherein P_3 is a residue selected from the group consisting of Phe, Glu, 15 and Pro; P_2 is any amino acid residue except Ser and P_2 is not Leu335 and is not Asn1720; and P_1 is Arg. The above modifications do not occur at a site of Factor VIII:C activation. Additionally, the non-activating Arg residues, Arg336 and Arg1719 can be substituted with 20 amino acids Pro, Glu, Asp, Asn, Gln, Ser and Tyr and/or the non-activating Arg residue Arg1721 can be substituted with Glu, Asp, Asn, Gln, Ser and Tyr, to impart a Factor VIII:C analog with improved properties.

25 **Definitions**

The term "Factor VIII:C polypeptide" refers to a polymer of amino acids and does not refer to a specific length of the product. Thus, peptides, oligopeptides, and proteins are included within the definition of 30 polypeptide. This term also does not exclude post-expression modifications of the polypeptide, for example, glycosylations, acetylations, phosphorylations and the like. Included within the definition are, for example, polypeptides containing one or more analogs of an amino 35 acid, including, for example, unnatural amino acids, polypeptides with substituted linkages, as well as other modifications known in the art, both naturally occurring

and non-naturally occurring. A Factor VIII:C polypeptide includes but is not limited to, for example, the following Factor VIII:C polypeptides: (a) a full-length Factor VIII:C molecule comprising a signal peptide and 5 all A, B, and C domains; (b) a mature Factor VIII:C molecule comprising all A, B, and C domains and lacking the signal peptide; (c) a truncated Factor VIII:C molecule lacking the signal peptide and at least a portion of the B domain; (d) a cleaved Factor VIII:C 10 molecule comprising a light chain subunit of about 80 kD; (e) a cleaved Factor VIII:C molecule comprising a heavy chain fragment of about 90 kD; (f) a cleaved Factor VIII:C molecule comprising a heavy chain fragment of about 50 kD; (g) a cleaved Factor VIII:C molecule 15 comprising a heavy chain fragment of about 43 kD; and (h) a cleaved Factor VIII:C molecule comprising a light chain fragment of about 73 kD.

Factor VIII:C polypeptides also include muteins or derivatives of the polypeptides with conservative 20 amino acid changes that do not alter the biological activity of the polypeptide from which the mutein or derivative is made. Such muteins or derivatives may have, for example, amino acid insertions, deletions, or substitutions in the relevant molecule that do not 25 substantially affect its properties. For example, the mutein or derivative can include conservative amino acid substitutions, such as substitutions which preserve the general charge, hydrophobicity/hydrophilicity, and/or stearic bulk of the amino acid substituted, for example 30 Gly/Ala; Val/Ile/Leu; Asp/Glu; Lys/Arg; Asn/Gln; and Phe/Trp/Tyr. The mutein or derivative should exhibit the same general structure as the native polypeptide, and may also include polypeptides having one or more peptide mimics or peptoids.

35 The term "active" in reference to the polypeptide analogs herein refers to biological activity, such as coagulation or pro-coagulation activity. Such

activity is measured by using standard assays for blood plasma samples, such as, for example, the Coatest assay or the activated partial thromboplastin time test (APTT). An "active" Factor VIII:C polypeptide analog will have at 5 least about 50% of the coagulation or pro-coagulation activity displayed by the native molecule, preferably at least about 60% to 80%, and more preferably at least about 90% or more of the coagulation or procoagulation activity displayed by the native Factor VIII:C molecule.

10 A "nucleic acid molecule" as used herein, refers to either RNA or DNA or its complementary strands thereof, that contains a nucleotide sequence.

The term "regulatory element" refers to an expression control sequence that is conventionally used to effect expression of a gene. A regulatory element includes one or more components that affect transcription 5 or translation, including transcription and translation signals. Such a sequence can be derived from a natural source or synthetically made, as in hybrid promoters and includes, for example, one or more of a promoter sequence, an enhancer sequence, a combination 10 promoter/enhancer sequence, an upstream activation sequence, a downstream termination sequence, a polyadenylation sequence, an optimal 5' leader sequence to optimize initiation of translation, and a Shine-Dalgarno sequence. The expression control sequence that 15 is appropriate for expression of the present polypeptide differs depending upon the host system in which the polypeptide is to be expressed. For example, in prokaryotes, such a sequence can include one or more of a promoter sequence, a ribosomal binding site, and a 20 transcription termination sequence. In eukaryotes, for example, such a sequence can include one or more of a promoter sequence, and a transcription termination sequence. If any component that is necessary for transcription or translation is lacking in the nucleic 25 acid molecule of the present invention, such a component

can be supplied by a vector. Regulatory elements suitable for use herein may be derived from a prokaryotic source, an eukaryotic source, a virus or viral vector or from a linear or circular plasmid.

5 The term "regulatory control" refers to control of expression of a polynucleotide sequence by a regulatory element to which the polynucleotide sequence is operably linked. The nature of such regulatory control differs depending upon the host organism. In
10 prokaryotes, such regulatory control is effected by regulatory sequences which generally include, for example, a promoter, and/or a transcription termination sequence. In eukaryotes, generally, such regulatory sequences include, for example, a promoter and/or a
15 transcription termination sequence. Additionally, other components which control of expression, for example, a signal peptide sequence or a secretory leader sequence for secretion of the polypeptide, and a terminator for transcriptional termination, may be attached thereto to
20 facilitate regulatory control of expression.

By "therapeutically effective amount" is meant an amount of analog that will improve the blood coagulation properties as compared to coagulation in the absence of the analog. A "therapeutically effective amount" will fall within a relatively broad range that can be determined through routine trials. The activity of the Factor VIII:C analogs of the invention may be determined by means known in the art, for example, by using the commercially available Coatest assay.
30 Preferably, the effective amount is sufficient to bring about prevention of further deterioration or treatment to improve coagulation, that is, to enhance coagulation properties such that hemostasis is achieved. The exact amount necessary will vary depending on the subject being
35 treated; the age and general condition of the individual to be treated; the functionality of the endogenous Factor VIII:C gene present in the individual; and the mode of

administration, among other factors. An appropriate effective amount can be readily determined by one of skill in the art. For example, depending on the severity of active Factor VIII:C polypeptide deficiency, up to 5 about 1000 to about 3000 U of Factor VIII:C polypeptide analog can be given to an average person such as a 70 kg male patient. Alternatively, sufficient Factor VIII:C polypeptide analog or analog complex can be given to establish a plasma level of about 0.5 to about 2 U/ml of 10 Factor VIII:C analog or combination analog and native polypeptide. See U.S. Patent Nos. 3,631,018; 3,652,530, and 4,069,216 for methods of administration and amounts.

The term "pharmaceutically acceptable excipient" refers to an excipient for administration of a 15 therapeutic agent, *in vivo*, and refers to any pharmaceutical agent that does not itself induce the production of antibodies harmful to the individual receiving the composition, and which may be administered without undue toxicity. Pharmaceutically acceptable 20 carriers in therapeutic compositions may contain liquids such as water, saline, glycerol and ethanol. Pharmaceutically acceptable salts can be included therein, for example, mineral acid salts such as hydrochlorides, hydrobromides, phosphates, sulfates, and 25 the like; and the salts of organic acids such as acetates, propionates, malonates, benzoates, and the like. Additionally, auxiliary substances, such as wetting or emulsifying agents, pH buffering substances, and the like, may be present in such vehicles. Suitable 30 carriers may also be present and are generally large, slowly metabolized macromolecules such as proteins, polysaccharides, polylactic acids, polyglycolic acids, polymeric amino acids, amino acid copolymers, and inactive virus particles. Such carriers are well known 35 to those of ordinary skill in the art. A thorough discussion of pharmaceutically acceptable excipients is available in REMINGTON'S PHARMACEUTICAL SCIENCES (Mack

Pub. Co., N.J. 1991). Typically, the therapeutic compositions are prepared as injectables, either as liquid solutions or suspensions; solid forms suitable for solution in, or suspension in, liquid vehicles prior to 5 injection may also be prepared.

The present invention provides Factor VIII:C polypeptide analogs with improved properties. The analogs include Pro-Arg bonds or Arg-Pro bonds at non-activating Arg residues. Preferably, the modification 10 will not introduce an Arg-Ala, Arg-Met, Arg-Gly, Arg-Ser or Arg-Thr bond at the modified site.

Additionally, the modification can comprise creation of a tripeptide having the formula $P_3-P_2-P_1$, wherein P_3 is a residue selected from the group 15 consisting of Phe, Glu, and Pro; P_2 is any amino acid residue except Ser and P_2 is not Leu335 and is not Asn1720; and P_1 is Arg.

Thus, when the non-activating Arg residue is Arg220, representative modifications can comprise an 20 insertion of at least one Pro residue between Asp219 and Arg 220; an insertion of at least one Pro residue between Arg220 and Asp221; a deletion comprising residues Asp221, Ala222, Ala223, Ser224, Ala225, Arg226, Ala227, and Trp228; an insertion of at least one Phe residue between 25 Gln218 and Asp219; an insertion of at least one Glu residue between Gln218 and Asp219; an insertion of at least one Pro residue between Gln218 and Asp219; a deletion comprising residues Thr212, Lys213, Asn214, Ser215, Leu216, Met217, and Gln218; a substitution of at 30 least one Phe residue at Gln218; a substitution of at least one Glu residue at Gln218; and/or a substitution of at least one Pro residue at Gln218.

When the non-activating Arg residue is Arg226, representative modifications can comprise an insertion of 35 at least one Pro residue between Ala225 and Arg226; an insertion of at least one Pro residue between Arg226 and Ala227; a deletion comprising residues Ala227 and Trp228;

a deletion comprising residues Ala227 and Trp228 and insertion of at least one Pro residue to replace the deleted residues; a deletion comprising residues Ala227, Trp228, Pro229, and Lys230, and an insertion of at least 5 one Pro residue to replace the deleted residues; an insertion of at least one Phe residue between Ser224 and Ala225; an insertion of at least one Glu residue between Ser224 and Ala225; an insertion of at least one Pro residue between Ser224 and Ala225; (d) a substitution of 10 at least one Phe residue at Ser224; a substitution of at least one Glu residue at Ser224; and/or a substitution of at least one Pro residue at Ser224.

When the non-activating Arg residue is Arg250, representative modifications can comprise an insertion of 15 at least one Pro residue between His249 and Arg250; an insertion of at least one Pro residue between Arg250 and Lys251; a deletion comprising residues Lys251 and Ser252 and insertion of at least one Pro residue to replace the deleted residues; a substitution of Lys251 with at least 20 one Pro residue; a deletion comprising residues Gly244, Leu245, Ile246, Gly247, Cys248, and His249; a deletion comprising residues Arg240, Ser241, Leu242, Pro243, Gly244, Leu245, Ile246, Gly247, Cys248, and His249 and an insertion of at least one Pro residue to replace the 25 deleted residues; an insertion of at least one Phe residue between Cys248 and His249; an insertion of at least one Glu residue between Cys248 and His249; an insertion of at least one Pro residue between Cys248 and His249; a deletion comprising residues Gly244, Leu245, 30 Ile246, Gly247, and Cys248; a substitution of at least one Phe residue at Cys248; a substitution of at least one Glu residue at Cys248; and/or a substitution of at least one Pro residue at Cys248.

When the non-activating Arg residue is Arg279, 35 representative modifications can comprise an insertion of at least one Pro residue between Val278 and Arg279; an insertion of at least one Pro residue between Arg279 and

Asn280; a deletion comprising residues Asn280, His281, and Arg282 and an insertion of at least one Pro residue to replace the deleted residues; a deletion comprising Asn280, His281, Arg282, Gln283, Ala284, and Ser285, and 5 an insertion of at least one Pro residue to replace the deleted residues; an insertion of at least one Phe residue between Leu277 and Val278; an insertion of at least one Glu residue between Leu277 and Val278; an insertion of at least one Pro residue between Leu277 and 10 Val278; a deletion comprising residue Leu277; a deletion comprising residue Val278; a deletion comprising residues Gly273, His274, Thr275, Phe276, and Leu277; a deletion comprising residues Leu271, Glu272, Gly273, His274, Thr275, Phe276, and Leu277; a substitution of at least 15 one Phe residue at Leu277; a substitution of at least one Glu residue at Leu277; and/or a substitution of at least one Pro residue at Leu277.

When the non-activating Arg residue is Arg282, representative modifications can comprise an insertion of 20 at least one Pro residue between His281 and Arg282; an insertion of at least one Pro residue between Arg282 and Gln283; a deletion comprising residues Arg279, Asn280, and His281 and an insertion of at least one Pro residue to replace the deleted residues; a deletion comprising 25 residues Gln283, Ala284, and Ser285, and an insertion of at least one Pro residue to replace the deleted residues; a deletion comprising residues Gln283, Ala284, Ser285, Leu286, Glu287, Ile288, and Ser289; an insertion of at least one Phe residue between Asn280 and His281; an 30 insertion of at least one Glu residue between Asn280 and His281; an insertion of at least one Pro residue between Asn280 and His281; a deletion comprising residues Leu277, Val278, Arg279, and Asn280; a deletion comprising residues Gly273, His274, Thr275, Phe276, Leu277, Val278, 35 Arg279, and Asn280; a substitution of at least one Phe residue at Asn280; a substitution of at least one Glu

residue at Asn280; and/or a substitution of at least one Pro residue at Asn280.

When the non-activating Arg residue is Arg336, representative modifications can comprise an insertion of 5 at least one Pro residue between Leu335 and Arg336; a deletion comprising residues Gln334, and Leu335; an insertion of at least one Pro residue between Arg336 and Met337; a deletion comprising residues Met337, and Lys338 and insertion of at least one Pro residue in place of the 10 deleted residues; a deletion comprising residue Leu335 and an insertion of at least one Phe residue between Pro333 and Gln334; a deletion comprising residue Leu335 and an insertion of at least one Glu residue between Pro333 and Gln334; a deletion comprising residue Leu335 15 and an insertion of at least one Pro residue between Pro333 and Gln334; a deletion comprising residue Leu335; a deletion comprising residues Gln334, and Leu335; a deletion comprising residues Pro333, Gln334, and Leu335; a deletion comprising residues Glu332, Pro333, Gln334, 20 and Leu335; a deletion comprising residue Leu335 and a substitution of at least one Phe residue at Pro333; and/or a deletion comprising residue Leu335 and a substitution of at least one Glu residue at Pro333.

When the non-activating Arg residue is Arg359, 25 representative modifications can comprise an insertion of at least one Pro residue between Val358 and Arg359; an insertion of at least one Pro residue between Arg359 and Phe360; a deletion comprising residues Phe360, Asp361, Asp362, Asp363, Asn364, and Ser365; a deletion comprising 30 residues Phe360, Asp361, Asp362, Asp363, Asn364, Ser365, Pro366, and Ser367, and an insertion of at least one Pro residue to replace the deleted residues; an insertion of at least one Phe residue between Val357 and Val358; an insertion of at least one Glu residue between Val357 and 35 Val358; an insertion of at least one Pro residue between Val357 and Val358; a deletion comprising residues Met355, Asp356, and Val357; a substitution of at least one Phe

residue at Val357; a substitution of at least one Glu residue at Val357; and/or a substitution of at least one Pro residue at Val357.

When the non-activating Arg residue is Arg562, 5 representative modifications can comprise an insertion of at least one Pro residue between Gln561 and Arg562; an insertion of at least one Pro residue between Arg562 and Gly563; a deletion comprising residues Ser558, Val559, Asp560, Gln561, and an insertion of at least one Pro 10 residue to replace the deleted residues; a deletion comprising residues Lys556, Glu557, Ser558, Val559, Asp560, Gln561, and an insertion of at least one Pro residue to replace the deleted residues; a deletion comprising residues Gly563, Asn564, Gln565, Ile566, 15 Met567, and Ser568, and an insertion of at least one Pro residue to replace the deleted residues; a deletion comprising residues Gly563, Asn564, Gln565, Ile566, Met567, Ser568, Asp569, Lys570, and Arg571, and an insertion of at least one Pro residue to replace the 20 deleted residues; an insertion of at least one Phe residue between Asp560 and Gln561; an insertion of at least one Glu residue between Asp560 and Gln561; an insertion of at least one Pro residue between Asp560 and Gln561; a deletion comprising residues Ser558, Val559, 25 and Asp560; a substitution of at least one Phe residue at Asp560; a substitution of at least one Glu residue at Asp560; and/or a substitution of at least one Pro residue at Asp560.

When the non-activating Arg residue is Arg747, 30 representative modifications can comprise an insertion of at least one Pro residue between Ser746 and Arg747; a deletion comprising residue Ser746 and an insertion of at least one Pro residue to replace the deleted residue; a deletion comprising residue His748; a deletion comprising 35 residue His748 and an insertion of at least one Pro residue to replace the deleted residue; an insertion of at least one Pro residue between Arg747 and His748; a

deletion comprising residues His748, Pro749, and Ser750 and an insertion of at least one Pro residue to replace the deleted residues; a deletion comprising Ser743, Gln744, Asn745, Ser746; a deletion comprising His748, 5 Pro749, Ser750, Thr751, Arg752, Gln753, and Lys754, and an insertion of at least one Pro residue to replace the deleted residues; a deletion comprising residue Ser746 and an insertion of at least one Phe residue between Gln744 and Asn745; a deletion comprising residue Ser746 10 and an insertion of at least one Glu residue between Gln744 and Asn745; a deletion comprising residue Ser746 and an insertion of at least one Pro residue between Gln744 and Asn745; a deletion comprising residue Ser746 and a substitution of at least one Phe residue at Gln744; 15 a deletion comprising residue Ser746 and a substitution of at least one Glu residue at Gln744; and/or a deletion comprising residue Ser746 and a substitution of at least one Pro residue at Gln744.

When the non-activating Arg residue is Arg776, 20 representative modifications can comprise an insertion of at least one Pro residue between His775 and Arg776; an insertion of at least one Pro residue between Arg776 and Thr777; a deletion comprising residue Thr777; a deletion comprising residue Thr777 and an insertion of at least 25 one Pro residue to replace the deleted residue; a deletion comprising residues Trp772, Phe773, Ala774, and His775; a deletion comprising residues Trp772, Phe773, Ala774, and His775, and an insertion of at least one Pro residue to replace the deleted residues; a deletion 30 comprising residues Lys768, Thr769, Asp770, Pro771, Trp772, Phe773, Ala774, and His775, and an insertion of at least one Pro residue to replace the deleted residues; a deletion comprising residues Thr777, Pro778, met779, Pro780, and Lys781, and an insertion of at least one Pro 35 residue to replace the deleted residues; an insertion of at least one Phe residue between Ala774 and His775; an insertion of at least one Glu residue between Ala774 and

His775; an insertion of at least one Pro residue between Ala774 and His775; a deletion comprising residue Ala774; a deletion comprising residues Trp772, Phe773 and Ala774; a deletion comprising residues Lys768, Thr769, Asp770, 5 Pro771, Trp772, and Phe773; a substitution of at least one Phe residue at Ala774; a substitution of at least one Glu residue at Ala774; and/or a substitution of at least one Pro residue at Ala774.

When the non-activating Arg residue is Arg1310, 10 representative modifications can comprise an insertion of at least one Pro residue between Gln1309 and Arg1310; an insertion of at least one Pro residue between Arg1310 and Ser1311; a deletion comprising residue Ser1311 and an insertion of at least one Pro residue to replace the 15 residue; a deletion comprising residues Ser1311, Lys1312, and Arg1313 and an insertion of at least one Pro residue to replace the deleted residues; a deletion comprising residues Ser1311, Lys1312, Arg1313, Ala1314, Leu1315, and Lys1316, and an insertion of at least one Pro residue to 20 replace the deleted residues; a deletion comprising residues Ser1311, Lys1312, Arg1313, Ala1314, Leu1315, Lys1316, Gln1317, Phe1318, Arg1319, and Leu1320; a deletion comprising residues Ser1311, Lys1312, Arg1313, Ala1314, Leu1315, Lys1316, Gln1317, Phe1318, Arg1319, and 25 Leu1320, and an insertion of at least one Pro residue to replace the deleted residues; an insertion of at least one Phe residue between Thr1308 and Gln1309; an insertion of at least one Glu residue between Thr1308 and Gln1309; an insertion of at least one Pro residue between Thr1308 30 and Gln1309; a deletion comprising residues Val1307, and Thr1308; a substitution of at least one Phe residue at Thr1308; a substitution of at least one Glu residue at Thr1308; and/or a substitution of at least one Pro residue at Thr1308.

35 When the non-activating Arg residue is Arg1313, representative modifications can comprise an insertion of at least one Pro residue between Lys1312 and Arg1313; a

deletion comprising residue Lys1312 and an insertion of at least one Pro residue to replace the deleted residue; a deletion comprising residues Arg1310, Ser1311, and Lys1312, and an insertion of at least one Pro residue to replace the deleted residues; an insertion of at least one Pro residue between Arg1313 and Ala1314; a deletion comprising residues Ala1314, Leu1315, and Lys1316, and an insertion of at least one Pro residue to replace the deleted residues; a deletion comprising residues Ala1314, 5 Leu1315, Lys1316, Gln1317, Phe1318, and Arg1319 and an insertion of at least one Pro residue to replace the deleted residues; a deletion comprising residues Ala1314, Leu1315, Lys1316, Gln1317, Phe1318, Arg1319, and Leu1320; an insertion of at least one Phe residue between Ser1311 10 and Lys1312; an insertion of at least one Glu residue between Ser1311 and Lys1312; an insertion of at least one Pro residue between Ser1311 and Lys1312; a deletion comprising residues Val1307, Thr1308, Gln1309, Arg1310, and Ser1311; a substitution of at least one Phe residue 15 at Ser1311; a substitution of at least one Glu residue at Ser 1311; and/or a substitution of at least one Pro residue at Ser 1311.

When the non-activating Arg residue is Arg1645, representative modifications can comprise a deletion 25 comprising residues Val1642, Leu1643, and Lys1644; a deletion comprising residue Lys1644 and an insertion of at least one Pro residue to replace the deleted residue; an insertion of at least one Pro residue between Arg1645 and Lys1644; an insertion of at least one Pro residue 30 between Arg1645 and His1646; a deletion comprising residues His1646, Gln1647, and Arg1648 and an insertion of at least one Pro residue to replace the deleted residues; a deletion comprising residues His1646, Gln1647, Arg1648, Glu1649, Ile1650, Thr1651, and Arg1652, 35 and an insertion of at least one Pro residue to replace the deleted residues; an insertion of at least one Phe residue between Leu1643 and Lys1644; an insertion of at

least one Glu residue between Leu1643 and Lys1644; an insertion of at least one Pro residue between Leu1643 and Lys1644; a deletion comprising residues Val1642, and Leu1643; a deletion comprising residues Pro1641, Val1642, 5 and Leu1643; a substitution of at least one Phe residue at Leu1643; a substitution of at least one Glu residue at Leu1643; and/or a substitution of at least one Pro residue at Leu1643.

When the non-activating Arg residue is Arg1648, 10 representative modifications include a deletion comprising residues Val1642, Leu1643, Lys1644, Arg1645, His1646, and Gln1647; an insertion of at least one Pro residue between Gln1647 and Arg1648; a deletion comprising residues Lys1644, Arg1645, His1646, and 15 Gln1647 and an insertion of at least one Pro residue to replace the deleted residues; a deletion comprising residues Glu1649, Ile1650, Thr1651, and Arg1652 and an insertion of at least one Pro residue to replace the deleted residues; an insertion of at least one Pro 20 residue between Arg1648 and Glu1649; an insertion of at least one Phe residue between His1646 and Gln1647; an insertion of at least one Glu residue between His1646 and Gln1647; an insertion of at least one Pro residue between His1646 and Gln1647; a deletion comprising residues 25 Val1642, Leu1643, Lys1644, Arg1645, and His1646; a deletion comprising residues Pro1641, Val1642, Leu1643, Lys1644, Arg1645, and His1646; a substitution of at least one Phe residue at His1646; (g) a substitution of at least one Glu residue at His1646; and/or a substitution 30 of at least one Pro residue at His1646.

When the non-activating Arg residue is Arg1719, representative modifications a deletion comprising residues His1716, Val1717, and Leu1718; an insertion of at least one Pro residue between Leu1718 and Arg1719; a 35 deletion comprising residues Asn1720, and Arg1721 and an insertion of at least one Pro residue to replace the deleted residues; an insertion of at least one Pro

residue between Arg1719 and Asn1720; an insertion of at least one Phe residue between Val1717 and Leu1718; an insertion of at least one Glu residue between Val1717 and Leu1718; an insertion of at least one Pro residue between 5 Val1717 and Leu1718; a deletion comprising residues His1716, and Val1717; a substitution of at least one Phe residue at Val1717; a substitution of at least one Glu residue at Val1717; and/or a substitution of at least one Pro residue at Val1717.

10 When the non-activating Arg residue is Arg1721, representative modifications include a deletion comprising residues His1716, Val1717, Leu1718, Arg1719, and Asn1720; an insertion of at least one Pro residue between Asn1720 and Arg1721; an insertion of at least one 15 Pro residue between Arg1721 and Ala1722; a deletion comprising residues Ala1722, Gln1723, and Ser1724 and an insertion of at least one Pro residue to replace the deleted residues; a deletion comprising residues Ala1722, Gln1723, Ser1724, Gly1725, Ser1726, and Val1727; a 20 deletion comprising residues Ala1722, Gln1723, Ser1724, Gly1725, and Ser1726, and an insertion of at least one Pro residue to replace the deleted residues; a deletion comprising residue Asn1720 and an insertion of at least one Phe residue between Leu1718 and Arg1719; a deletion 25 comprising residue Asn1720 and an insertion of at least one Glu residue between Leu1718 and Arg1719; a deletion comprising residue Asn1720 and an insertion of at least one Pro residue between Leu1718 and Arg1719; a deletion comprising residues Val1717, Leu1718, Arg1719, and 30 Asn1720; a deletion comprising residue Asn1720 and a substitution of at least one Phe residue at Leu1718; a deletion comprising residue Asn1720 and a substitution of at least one Glu residue at Leu1718; and/or a deletion comprising residue Asn1720 and a substitution of at least 35 one Pro residue at Leu1718.

Other polypeptide analogs contemplated by the present invention are those analogs including

substitutions of the non-activating Arg residues found at positions Arg336, Arg1719 and Arg1721, numbered with respect to the native Factor VIII:C polypeptide sequence. For example, based on the model of Factor VIII:C 5 described in Pan et al., *Nature Structural Biology* (1995) 2:740-744, any of amino acids Pro, Glu, Asp, Asn, Gln, Ser and Tyr can be substituted at position Arg336 and/or position Arg1719, to generate a Factor VIII:C polypeptide analog with improved properties. Similarly, any of amino 10 acids Glu, Asp, Asn, Gln, Ser and Tyr can be substituted at position Arg1721. Preferably, the Factor VIII:C polypeptide analog of this embodiment will include a substitution of Arg336 with Pro, a substitution of Arg 1719 with Pro and/or a substitution of Arg1721 with Glu.

15 Nucleic acid molecules encoding the present Factor VIII:C polypeptide analogs can be made by modifying the native nucleic acid sequences that encode the Factor VIII:C polypeptide or cDNA sequences that encode the Factor VIII:C polypeptides. Such modification 20 can be done by conventional techniques such as site-directed mutagenesis. For example, the M13 method for site directed mutagenesis is known, as described in Zoller and Smith, *Nucleic Acids Res.* (1982) 10: 6487-6500, *Methods Enzymol.* (1983) 100: 468-500, and *DNA* 25 (1984) 3: 479-488, using single stranded DNA, and the method of Morinaga et al. *Bio/technol.* 636-639 (July 1984), using heteroduplexed DNA. According to the method of the invention, by site-directed mutagenesis, one or more of the codons encoding a residue adjacent to an 30 arginine, preferably, at the carboxy side, can be mutated by substitution, deletion or addition, to a codon encoding proline, thereby creating polypeptides with either Pro-Arg bonds or Arg-Pro bonds where none existed before, provided that the modification does not affect an 35 activation cleavage site, such as Arg372 and Arg740. The codons encoding proline include CCU, CCT, CCG, CCA, and CCC. A description of a protocol suitable for use herein

for mutagenesis of specific sites of a Factor VIII:C expression plasmid can be found in WO 87/07144.

The nucleic acid molecules of the present invention can also be made synthetically by piecing together nucleic acid molecules encoding heavy and light chain fragments derived from cDNA clones or genomic clones containing Factor VIII:C coding sequences, preferably cDNA clones, using known linker sequences. Alternatively, the entire sequence or portions of nucleic acid sequences encoding analogs described above may be prepared by synthetic methods (e.g. using DNA synthesis machines). Once made, the nucleic acid molecules can be inserted in vectors for production of recombinant vectors for transcription and translation of the nucleic acid molecules.

One skilled in the art of DNA cloning and in possession of the DNA encoding native Factor VIII:C polypeptide will be able to prepare suitable DNA molecules for production of the present analogs using known cloning procedures (e.g. restriction enzyme digestion, exonuclease digestion, ligation, and other appropriate procedures) outlined in any of the following: Sambrook, et al, MOLECULAR CLONING: A LABORATORY MANUAL 2nd ed. (Cold Spring Harbor Laboratory Press, 1989); DNA CLONING, Vol. I and II, D.N. Glover ed. (IRL Press, 1985); OLIGONUCLEOTIDE SYNTHESIS, M.J. Gait ed. (IRL Press, 1984); NUCLEIC ACID HYBRIDIZATION, B.D. Hames & S.J. Higgins eds. (IRL Press, 1984); TRANSCRIPTION AND TRANSLATION, B.D. Hames & S.J. Higgins eds., (IRL Press, 1984); ANIMAL CELL CULTURE, R.I. Freshney ed. (IRL Press, 1986); IMMOBILIZED CELLS AND ENZYMES, K. Mosbach (IRL Press, 1986); B. Perbal, A PRACTICAL GUIDE TO MOLECULAR CLONING, Wiley (1984); the series, METHODS IN ENZYMOLOGY, Academic Press, Inc.; GENE TRANSFER VECTORS FOR MAMMALIAN CELLS, J.H. Miller and M.P. Calos eds. (Cold Spring Harbor Laboratory, 1987); METHODS IN ENZYMOLOGY, Vol. 154 and 155, Wu and Grossman, eds., and Wu, ed., respectively

(Academic Press, 1987), IMMUNOCHEMICAL METHODS IN CELL AND MOLECULAR BIOLOGY, R.J. Mayer and J.H. Walker, eds. (Academic Press London, Harcourt Brace U.S., 1987), PROTEIN PURIFICATION: PRINCIPLES AND PRACTICE, 2nd ed.

5 (Springer-Verlag, N.Y. (1987), and HANDBOOK OF EXPERIMENTAL IMMUNOLOGY, Vol. I-IV, D.M. Weir et al, (Blackwell Scientific Publications, 1986); Kitts et al, Biotechniques, (1993), 14:810-817; Munemitsu et al, Mol. Cell. Biol., (1990) 10:5977-5982. Finally, a preferred 10 method of preparing nucleic acid molecules encoding the described analogs is by use of PCR techniques, especially overlapping PCR, as described in PCR PROTOCOLS: A GUIDE TO METHODS AND APPLICATIONS, Innis, Gelfand, Sninsky, and White (eds.) (Academic Press, 1990).

15 A vector suitable for use herein for the production of a recombinant vector comprises a nucleic acid sequence with one or more restriction enzyme recognition sites into which the present nucleic acid molecule of the invention can be inserted. This vector 20 also typically contains a selection marker for detection of the presence of the vector in the host cell. The vector can also provide, if desired, one or more regulatory elements or control sequences for expression of the nucleic acid molecule. For example, the present 25 vector can be derived from a plasmid, a virus, a cosmid, or a bacteriophage. This vector is typically capable of behaving as an autonomous unit of replication when introduced into a host cell. Moreover, the vector may be one that is capable of episomal existence or of 30 integration into the host cell genome. A wide variety of replication systems are available, typically derived from viruses that infect mammalian host cells. Illustrative replication systems include the replication systems from Simian virus 40, adenovirus, bovine papilloma virus, 35 polyoma virus, Epstein Barr virus, and the like. Thus, the nucleic acid molecule of the present invention can be inserted at an appropriate restriction site in the vector

so as to be placed under the control of one or more regulatory elements in the vector to form a recombinant vector that can be used for transfection or transformation of a host cell.

5 The host cells of the invention can be, for example, prokaryotic or eukaryotic host cells, including bacterial, yeast, insect and mammalian expression systems. Preferably the analogs of the present invention are expressed in mammalian host cell systems.

10 The regulatory elements to be used in the vector depend on the host system that is to be utilized. For example, a prokaryotic host cell can be used for amplification of the nucleic acid molecule of the present invention, while an eukaryotic host cell can be used for 15 expression of the Factor VIII:C polypeptide analogs.

The expression cassettes are introduced into the host cell by conventional methods, depending on the expression system used, as described further below.

Where viruses are involved, transfection or transduction 20 may be employed. The particular manner in which the host cell is transformed is not critical to this invention, depending substantially upon whether the expression cassettes are joined to a replication system and the nature of the replication system and associated genes.

25 Coexpression of more than one Factor VIII:C polypeptide analog may be desired. For example, it may be desirable to express the light and heavy chains using separate constructs. In this regard, either or both of the light and heavy chains may include modifications as 30 described above. "Coexpression" as used herein refers to the expression of two or more Factor VIII:C polypeptides in a single host cell. Thus, for example, the expression of the 90 kD species and the 80 kD species in a single host cell, would constitute "coexpression" as used 35 herein. The polynucleotides encoding for the polypeptides can be harbored in a single vector, either under the control of the same regulatory elements or

under the control of separate elements. Thus, the production of a fusion protein including active portions of the two or more Factor VIII:C polypeptides would be considered "coexpressed" for purposes of the present 5 definition as would the expression of two genes as a dicistronic construct employing an internal ribosome entry site. Similarly, proteins expressed from the same vector but driven by separate regulatory elements, would also be considered "coexpressed." The term also refers 10 to the expression of two or more proteins from separate constructs. Thus, the expression of proteins encoded from genes present on separate vectors in a host cell would also be considered "coexpression" for purposes of the present invention.

15 The transformed/transfected cells are then grown in an appropriate nutrient medium. If separate constructs encoding heavy and light chain analogs have been used for coexpression, the product can be obtained as a complex of the two Factor VIII:C chains, so that the 20 media or cell lysate may be isolated and the Factor VIII:C active complex extracted and purified. Similarly, the full-length molecule can be isolated and treated under complex-forming conditions, e.g., with the addition of calcium and the appropriate enzymes, to form the 25 active complex. Various means are available for extraction and purification, such as affinity chromatography, ion exchange chromatography, hydrophobic chromatography, electrophoresis, solvent-solvent extraction, selective precipitation, and the like. The 30 particular manner in which the product is isolated is not critical to this invention, and is selected to minimize denaturation or inactivation and maximize the isolation of a high-purity active product.

Expression in Bacterial Cells

Bacterial expression systems can be used to produce the subject Factor VIII:C polypeptide analogs and nucleic acid sequences encoding the analogs. Control 5 elements for use in bacterial systems include promoters, optionally containing operator sequences, and ribosome binding sites. Useful promoters include sequences derived from sugar metabolizing enzymes, such as galactose, lactose (*lac*) and maltose. Additional 10 examples include promoter sequences derived from biosynthetic enzymes such as tryptophan (*trp*), the β -lactamase (*bla*) promoter system, bacteriophage λ PL, and T7. In addition, synthetic promoters can be used, such as the *tac* promoter. The β -lactamase and lactose 15 promoter systems are described in Chang et al., *Nature* (1978) 275: 615, and Goeddel et al., *Nature* (1979) 281: 544; the alkaline phosphatase, tryptophan (*trp*) promoter system are described in Goeddel et al., *Nucleic Acids Res.* (1980) 8: 4057 and EP 36,776 and hybrid promoters 20 such as the *tac* promoter is described in U.S. Patent No. 4,551,433 and deBoer et al., *Proc. Natl. Acad. Sci. USA* (1983) 80: 21-25. However, other known bacterial promoters useful for expression of eukaryotic proteins are also suitable. A person skilled in the art would be 25 able to operably ligate such promoters to the present Factor VIII:C polypeptide analog coding sequences, for example, as described in Siebenlist et al., *Cell* (1980) 20: 269, using linkers or adapters to supply any required restriction sites. Promoters for use in bacterial 30 systems also generally will contain a Shine-Dalgarno (SD) sequence operably linked to the DNA encoding the Factor VIII:C analog polypeptide. For prokaryotic host cells that do not recognize and process the native polypeptide signal sequence, the signal sequence can be substituted 35 by a prokaryotic signal sequence selected, for example, from the group of the alkaline phosphatase, penicillinase, Ipp, or heat stable enterotoxin II

leaders. The origin of replication from the plasmid pBR322 is suitable for most Gram-negative bacteria.

The foregoing systems are particularly compatible with *Escherichia coli*. However, numerous 5 other systems for use in bacterial hosts including Gram-negative or Gram-positive organisms such as *Bacillus spp.*, *Streptococcus spp.*, *Streptomyces spp.*, *Pseudomonas* species such as *P. aeruginosa*, *Salmonella typhimurium*, or *Serratia marcescans*, among others. Methods for 10 introducing exogenous DNA into these hosts typically include the use of CaCl_2 or other agents, such as divalent cations and DMSO. DNA can also be introduced into bacterial cells by electroporation, nuclear injection, or protoplast fusion as described generally in 15 Sambrook et al. (1989), cited above. These examples are illustrative rather than limiting. Preferably, the host cell should secrete minimal amounts of proteolytic enzymes. Alternatively, *in vitro* methods of cloning, e.g., PCR or other nucleic acid polymerase reactions, are 20 suitable.

Prokaryotic cells used to produce the Factor VIII:C analog polypeptides of this invention are cultured in suitable media, as described generally in Sambrook et al., cited above.

25

Expression in Yeast Cells

Yeast expression systems can also be used to produce the subject Factor VIII:C polypeptide analogs and nucleic acid sequences encoding the analogs. Expression 30 and transformation vectors, either extrachromosomal replicons or integrating vectors, have been developed for transformation into many yeasts. For example, expression vectors have been developed for, among others, the following yeasts: *Saccharomyces cerevisiae*, as described 35 in Hinnen et al., *Proc. Natl. Acad. Sci. USA* (1978) 75: 1929; Ito et al., *J. Bacteriol.* (1983) 153: 163; *Candida albicans* as described in Kurtz et al., *Mol. Cell. Biol.*

(1986) 6: 142; *Candida maltosa*, as described in Kunze et al., *J. Basic Microbiol.* (1985) 25: 141; *Hansenula polymorpha*, as described in Gleeson et al., *J. Gen. Microbiol.* (1986) 132: 3459 and Roggenkamp et al., *Mol. Gen. Genet.* (1986) 202: 302); *Kluyveromyces fragilis*, as described in Das et al., *J. Bacteriol.* (1984) 158: 1165; *Kluyveromyces lactis*, as described in De Louvencourt et al., *J. Bacteriol.* (1983) 154: 737 and Van den Berg et al., *Bio/Technology* (1990) 8: 135; *Pichia guillerimondii*, 10 as described in Kunze et al., *J. Basic Microbiol.* (1985) 25: 141; *Pichia pastoris*, as described in Cregg et al., *Mol. Cell. Biol.* (1985) 5: 3376 and U.S. Patent Nos. 4,837,148 and 4,929,555; *Schizosaccharomyces pombe*, as described in Beach and Nurse, *Nature* (1981) 300: 706; and 15 *Yarrowia lipolytica*, as described in Davidow et al., *Curr. Genet.* (1985) 10: 380 and Gaillardin et al., *Curr. Genet.* (1985) 10: 49, *Aspergillus* hosts such as *A. nidulans*, as described in Ballance et al., *Biochem. Biophys. Res. Commun.* (1983) 112: 284-289; Tilburn et 20 al., *Gene* (1983) 26: 205-221 and Yelton et al., *Proc. Natl. Acad. Sci. USA* (1984) 81: 1470-1474, and *A. niger*, as described in Kelly and Hynes, *EMBO J.* (1985) 4: 475479; *Trichoderma reesia*, as described in EP 244,234, and filamentous fungi such as, e.g., *Neurospora*, 25 *Penicillium*, *Tolypocladium*, as described in WO 91/00357.

Control sequences for yeast vectors are known and include promoter regions from genes such as alcohol dehydrogenase (ADH), as described in EP 284,044, enolase, glucokinase, glucose-6-phosphate isomerase, 30 glyceraldehyde-3-phosphate-dehydrogenase (GAP or GAPDH), hexokinase, phosphofructokinase, 3-phosphoglycerate mutase, and pyruvate kinase (PyK), as described in EP 329,203. The yeast *PHO5* gene, encoding acid phosphatase, also provides useful promoter sequences, as described in 35 Myanohara et al., *Proc. Natl. Acad. Sci. USA* (1983) 80:1. Other suitable promoter sequences for use with yeast hosts include the promoters for 3-phosphoglycerate

kinase, as described in Hitzeman et al., *J. Biol. Chem.* (1980) 255: 2073, or other glycolytic enzymes, such as pyruvate decarboxylase, triosephosphate isomerase, and phosphoglucose isomerase, as described in Hess et al., *J. 5 Adv. Enzyme Reg.* (1968) 7: 149 and Holland et al., *Biochemistry* (1978) 17: 4900. Inducible yeast promoters having the additional advantage of transcription controlled by growth conditions, include from the list above and others the promoter regions for alcohol 10 dehydrogenase 2, isocytchrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, metallothionein, glyceraldehyde-3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose utilization. Suitable vectors and promoters 15 for use in yeast expression are further described in Hitzeman, EP 073,657. Yeast enhancers also are advantageously used with yeast promoters. In addition, synthetic promoters which do not occur in nature also function as yeast promoters. For example, upstream 20 activating sequences (UAS) of one yeast promoter may be joined with the transcription activation region of another yeast promoter, creating a synthetic hybrid promoter. Examples of such hybrid promoters include the ADH regulatory sequence linked to the GAP transcription 25 activation region, as described in U.S. Patent Nos. 4,876,197 and 4,880,734. Other examples of hybrid promoters include promoters which consist of the regulatory sequences of either the *ADH2*, *GAL4*, *GAL10*, or *PHO5* genes, combined with the transcriptional activation 30 region of a glycolytic enzyme gene such as *GAP* or *PyK*, as described in EP 164,556. Furthermore, a yeast promoter can include naturally occurring promoters of non-yeast origin that have the ability to bind yeast RNA polymerase and initiate transcription.

35 Other control elements which may be included in the yeast expression vectors are terminators, for example, from *GAPDH* and from the enolase gene, as

described in Holland et al., *J. Biol. Chem.* (1981) 256: 1385, and leader sequences which encode signal sequences for secretion. DNA encoding suitable signal sequences can be derived from genes for secreted yeast proteins, 5 such as the yeast invertase gene as described in EP 012,873 and JP 62,096,086 and the α -factor gene, as described in U.S. Patent Nos. 4,588,684, 4,546,083 and 4,870,008; EP 324,274; and WO 89/02463. Alternatively, leaders of non-yeast origin, such as an interferon 10 leader, also provide for secretion in yeast, as described in EP 060,057.

Methods of introducing exogenous DNA into yeast hosts are well known in the art, and typically include either the transformation of spheroplasts or of intact 15 yeast cells treated with alkali cations. Transformations into yeast can be carried out according to the method described in Van Solingen et al., *J. Bact.* (1977) 130: 946 and Hsiao et al., *Proc. Natl. Acad. Sci. USA* (1979) 76: 3829. However, other methods for introducing DNA into 20 cells such as by nuclear injection, electroporation, or protoplast fusion may also be used as described generally in Sambrook et al., cited above.

For yeast secretion the native polypeptide signal sequence may be substituted by the yeast 25 invertase, α -factor, or acid phosphatase leaders. The origin of replication from the 2μ plasmid origin is suitable for yeast. A suitable selection gene for use in yeast is the *trp1* gene present in the yeast plasmid described in Kingsman et al., *Gene* (1979) 7: 141 or 30 Tschemper et al., *Gene* (1980) 10: 157. The *trp1* gene provides a selection marker for a mutant strain of yeast lacking the ability to grow in tryptophan. Similarly, Leu2-deficient yeast strains (ATCC 20,622 or 38,626) are complemented by known plasmids bearing the *Leu2* Gene.

35 For intracellular production of the present polypeptides in yeast, a sequence encoding a yeast protein can be linked to a coding sequence of a Factor

VIII:C polypeptide analog to produce a fusion protein that can be cleaved intracellularly by the yeast cells upon expression. An example, of such a yeast leader sequence is the yeast ubiquitin gene.

5

Expression in Insect Cells

Insect expression systems can be used to produce the Factor VIII:C polypeptide analogs and nucleic acid sequences encoding the analogs. For example,

10 baculovirus expression vectors (BEVs) are recombinant insect viruses in which the coding sequence for a foreign gene to be expressed is inserted behind a baculovirus promoter in place of a viral gene, e.g., polyhedrin, as described in Smith and Summers, U.S. Pat. No., 4,745,051.

15 An expression construct herein includes a DNA vector useful as an intermediate for the infection or transformation of an insect cell system, the vector generally containing DNA coding for a baculovirus transcriptional promoter, optionally but preferably, 20 followed downstream by an insect signal DNA sequence capable of directing secretion of a desired protein, and a site for insertion of the foreign gene encoding the foreign protein, the signal DNA sequence and the foreign gene being placed under the transcriptional control of a 25 baculovirus promoter, the foreign gene herein being the coding sequence of a Factor VIII:C polypeptide analog of this invention.

The promoter for use herein can be a baculovirus transcriptional promoter region derived from 30 any of the over 500 baculoviruses generally infecting insects, such as, for example, the Orders Lepidoptera, Diptera, Orthoptera, Coleoptera and Hymenoptera including, for example, but not limited to the viral DNAs of *Autographa californica* MNPV, *Bombyx mori* NPV, 35 *rrichoplusia ni* MNPV, *Rachlplusia ou* MNPV or *Galleria mellonella* MNPV, *Aedes aegypti*, *Drosophila melanogaster*, *Spodoptera frugiperda*, and *Trichoplusia ni*. Thus, the

1 baculovirus transcriptional promoter can be, for example, a baculovirus immediate-early gene IEI or IEN promoter; an immediate-early gene in combination with a baculovirus delayed-early gene promoter region selected from the
5 group consisting of a 39K and a *Hind*III fragment containing a delayed-early gene; or a baculovirus late gene promoter. The immediate-early or delayed-early promoters can be enhanced with transcriptional enhancer elements.

10 Particularly suitable for use herein is the strong polyhedrin promoter of the baculovirus, which directs a high level of expression of a DNA insert, as described in Friesen et al. (1986) "The Regulation of Baculovirus Gene Expression" in: THE MOLECULAR BIOLOGY OF
15 BACULOVIRUSES (W.Doerfler, ed.); EP 127,839 and EP 155,476; and the promoter from the gene encoding the p10 protein, as described in Vlak et al., *J. Gen. Virol.* (1988) 69: 765-776. The plasmid for use herein usually also contains the polyhedrin polyadenylation
20 signal, as described in Miller et al., *Ann. Rev. Microbiol.* (1988) 42: 177 and a procaryotic ampicillin-resistance (amp) gene and an origin of replication for selection and propagation in *E. coli*. DNA encoding suitable signal sequences can also be included and is
25 generally derived from genes for secreted insect or baculovirus proteins, such as the baculovirus polyhedrin gene, as described in Carbonell et al., *Gene* (1988) 73: 409, as well as mammalian signal sequences such as those derived from genes encoding human α -interferon as
30 described in Maeda et al., *Nature* (1985) 315: 592-594; human gastrin-releasing peptide, as described in Lebacq-Verheyden et al., *Mol. Cell. Biol.* (1988) 8: 3129; human IL-2, as described in Smith et al., *Proc. Natl. Acad. Sci. USA* (1985) 82: 8404; mouse IL-3, as described in
35 Miyajima et al., *Gene* (1987) 58: 273; and human glucocerebrosidase, as described in Martin et al., *DNA* (1988) 7:99.

Numerous baculoviral strains and variants and corresponding permissive insect host cells from hosts such as *Spodoptera frugiperda* (caterpillar), *Aedes aegypti* (mosquito), *Aedes albopictus* (mosquito), 5 *Drosophila melanogaster* (fruitfly), and *Bombyx mori* host cells have been identified and can be used herein. See, for example, the description in Luckow et al., Bio/Technology(1988) 6: 47-55, Miller et al., in GENETIC ENGINEERING (Setlow, J.K. et al. eds.), Vol. 8 (Plenum 10 Publishing, 1986), pp. 277-279, and Maeda et al., *Nature*, (1985) 315: 592-594. A variety of such viral strains are publicly available, e.g., the L-1 variant of *Autographa californica* NPV and the Bm-5 strain of *Bombyx mori* NPV. Such viruses may be used as the virus for transfection of 15 host cells such as *Spodoptera frugiperda* cells.

Other baculovirus genes in addition to the polyhedrin promoter may be employed to advantage in a baculovirus expression system. These include immediate-early (alpha), delayed-early (beta), late 20 (gamma), or very late (delta), according to the phase of the viral infection during which they are expressed. The expression of these genes occurs sequentially, probably as the result of a "cascade" mechanism of transcriptional regulation. Thus, the immediate-early genes are 25 expressed immediately after infection, in the absence of other viral functions, and one or more of the resulting gene products induces transcription of the delayed-early genes. Some delayed-early gene products, in turn, induce transcription of late genes, and finally, the very late 30 genes are expressed under the control of previously expressed gene products from one or more of the earlier classes. One relatively well defined component of this regulatory cascade is IEI, a preferred immediate-early gene of *Autographa californica* nuclear polyhedrosis virus 35 (AcMNPV). IEI is pressed in the absence of other viral functions and encodes a product that stimulates the transcription of several genes of the delayed-early

class, including the preferred 39K gene, as described in Guarino and Summers, *J. Virol.* (1986) 57: 563-571 and *J. Virol.* (1987) 61: 2091-2099 as well as late genes, as described in Guanno and Summers, *Virol.* (1988) 162: 5 444-451.

Immediate-early genes as described above can be used in combination with a baculovirus gene promoter region of the delayed-early category. Unlike the immediate-early genes, such delayed-early genes require 10 the presence of other viral genes or gene products such as those of the immediate-early genes. The combination of immediate-early genes can be made with any of several delayed-early gene promoter regions such as 39K or one of the delayed-early gene promoters found on the HindIII 15 fragment of the baculovirus genome. In the present instance, the 39 K promoter region can be linked to the foreign gene to be expressed such that expression can be further controlled by the presence of IEI, as described in L. A. Guarino and Summers (1986a), cited above; 20 Guarino & Summers (1986b) *J. Virol.*, (1986) 60: 215-223, and Guarino et al. (1986c), *J. Virol.* (1986) 60: 224-229.

Additionally, when a combination of immediate-early genes with a delayed-early gene promoter region is used, enhancement of the expression of 25 heterologous genes can be realized by the presence of an enhancer sequence in direct cis linkage with the delayed-early gene promoter region. Such enhancer sequences are characterized by their enhancement of delayed-early gene expression in situations where the 30 immediate-early gene or its product is limited. For example, the hr5 enhancer sequence can be linked directly, in cis, to the delayed-early gene promoter region, 39K, thereby enhancing the expression of the cloned heterologous DNA as described in Guarino and 35 Summers (1986a), (1986b), and Guarino et al. (1986).

The polyhedrin gene is classified as a very late gene. Therefore, transcription from the polyhedrin

promoter requires the previous expression of an unknown, but probably large number of other viral and cellular gene products. Because of this delayed expression of the polyhedrin promoter, state-of-the-art BEVs, such as the 5 exemplary BEV system described by Smith and Summers in, for example, U.S. Pat. No., 4,745,051 will express foreign genes only as a result of gene expression from the rest of the viral genome, and only after the viral infection is well underway. This represents a limitation 10 to the use of existing BEVs. The ability of the host cell to process newly synthesized proteins decreases as the baculovirus infection progresses. Thus, gene expression from the polyhedrin promoter occurs at a time when the host cell's ability to process newly synthesized 15 proteins is potentially diminished for certain proteins such as human tissue plasminogen activator. As a consequence, the expression of secretory glycoproteins in BEV systems is complicated due to incomplete secretion of the cloned gene product, thereby trapping the cloned gene 20 product within the cell in an incompletely processed form.

While it has been recognized that an insect signal sequence can be used to express a foreign protein that can be cleaved to produce a mature protein, the 25 present invention is preferably practiced with a mammalian signal sequence for example the Factor VIII signal sequence.

An exemplary insect signal sequence suitable herein is the sequence encoding for a Lepidopteran 30 adipokinetic hormone (AKH) peptide. The AKH family consists of short blocked neuropeptides that regulate energy substrate mobilization and metabolism in insects. In a preferred embodiment, a DNA sequence coding for a Lepidopteran *Manduca sexta* AKH signal peptide can be 35 used. Other insect AKH signal peptides, such as those from the Orthoptera *Schistocerca gregaria* locus can also be employed to advantage. Another exemplary insect

signal sequence is the sequence coding for *Drosophila* cuticle proteins such as CPI, CP2, CP3 or CP4.

Currently, the most commonly used transfer vector that can be used herein for introducing foreign genes into AcNPV is pAc373. Many other vectors, known to those of skill in the art, can also be used herein. Materials and methods for baculovirus/insect cell expression systems are commercially available in a kit form from companies such as Invitrogen (San Diego CA) ("MaxBac" kit). The techniques utilized herein are generally known to those skilled in the art and are fully described in Summers and Smith, A MANUAL OF METHODS FOR BACULOVIRUS VECTORS AND INSECT CELL CULTURE PROCEDURES, Texas Agricultural Experiment Station Bulletin No. 1555, Texas A&M University (1987); Smith et al., *Mol. Cell. Biol.* (1983) 3: 2156, and Luckow and Summers (1989). These include, for example, the use of pVL985 which alters the polyhedrin start codon from ATG to ATT, and which introduces a *Bam*HI cloning site 32 basepairs downstream from the ATT, as described in Luckow and Summers, *Virology* (1989) 17:31.

Thus, for example, for insect cell expression of the present polypeptides, the desired DNA sequence can be inserted into the transfer vector, using known techniques. An insect cell host can be cotransformed with the transfer vector containing the inserted desired DNA together with the genomic DNA of wild type baculovirus, usually by cotransfection. The vector and viral genome are allowed to recombine resulting in a recombinant virus that can be easily identified and purified. The packaged recombinant virus can be used to infect insect host cells to express a Factor VIII:C polypeptide analog.

Other methods that are applicable herein are the standard methods of insect cell culture, cotransfection and preparation of plasmids are set forth in Summers and Smith (1987), cited above. This reference

also pertains to the standard methods of cloning genes into AcMNPV transfer vectors, plasmid DNA isolation, transferring genes into the AcMNPV genome, viral DNA purification, radiolabeling recombinant proteins and 5 preparation of insect cell culture media. The procedure for the cultivation of viruses and cells are described in Volkman and Summers, *J. Virol.* (1975) 19:820-832 and Volkman, et al., *J. Virol.* (1976) 19:820-832.

10 Expression in Mammalian Cells

Mammalian expression systems can also be used to produce the Factor VIII:C polypeptide analogs and nucleic acid sequences encoding the analogs. Typical promoters for mammalian cell expression include the SV40 15 early promoter, the CMV promoter, the mouse mammary tumor virus LTR promoter, the adenovirus major late promoter (Ad MLP), and the herpes simplex virus promoter, among others. Other non-viral promoters, such as a promoter derived from the murine metallothionein gene, will also 20 find use in mammalian constructs. Mammalian expression may be either constitutive or regulated (inducible), depending on the promoter. Typically, transcription termination and polyadenylation sequences will also be present, located 3' to the translation stop codon.

25 Preferably, a sequence for optimization of initiation of translation, located 5' to the Factor VIII:C polypeptide analog coding sequence, is also present. Examples of transcription terminator/polyadenylation signals include those derived from SV40, as described in Sambrook et al.

30 (1989) MOLECULAR CLONING: A LABORATORY MANUAL, 2d edition, (Cold Spring Harbor Press, Cold Spring Harbor, N.Y.). Introns, containing splice donor and acceptor sites, may also be designed into the constructs of the present invention.

35 Enhancer elements can also be used herein to increase expression levels of the mammalian constructs. Examples include the SV40 early gene enhancer, as

described in Dijkema et al., *EMBO J.* (1985) 4: 761 and the enhancer/promoter derived from the long terminal repeat (LTR) of the Rous Sarcoma Virus, as described in Gorman et al., *Proc. Natl. Acad. Sci. USA* (1982b) 79: 5 6777 and human cytomegalovirus, as described in Boshart et al., *Cell* (1985) 41: 521. A leader sequence can also be present which includes a sequence encoding a signal peptide, to provide for the secretion of the foreign protein in mammalian cells. Alternatively, the Factor 10 VIII signal peptide can be used. Preferably, there are processing sites encoded between the leader fragment and the gene of interest such that the leader sequence can be cleaved either *in vivo* or *in vitro*. The adenovirus 15 tripartite leader is an example of a leader sequence that provides for secretion of a foreign protein in mammalian cells.

There exist expression vectors that provide for the transient expression in mammalian cells of DNA encoding the Factor VIII:C analog polypeptides. In 20 general, transient expression involves the use of an expression vector that is able to replicate efficiently in a host cell, such that the host cell accumulates many copies of the expression vector and, in turn, synthesizes high levels of a desired polypeptide encoded by the 25 expression vector. Transient expression systems, comprising a suitable expression vector and a host cell, allow for the convenient positive identification of polypeptides encoded by cloned DNAs, as well as for the rapid screening of such polypeptides for desired 30 biological or physiological properties. Thus, transient expression systems are particularly useful for purposes of identifying additional polypeptides that have Factor VIII:C-like activity.

Once complete, the mammalian expression vectors 35 can be used to transform any of several mammalian cells. Methods for introduction of heterologous polynucleotides into mammalian cells are known in the art and include

dextran-mediated transfection, calcium phosphate precipitation, polybrene mediated transfection, protoplast fusion, electroporation, encapsulation of the polynucleotide(s) in liposomes, and direct microinjection 5 of the DNA into nuclei. General aspects of mammalian cell host system transformations have been described by Axel in U.S. 4,399,216. A synthetic lipid particularly useful for polynucleotide transfection is N-[1-(2,3-dioleyloxy)propyl]-N,N,N-trimethylammonium chloride, 10 which is commercially available under the name Lipofectin® (available from BRL, Gaithersburg, MD), and is described by Felgner et al., *Proc. Natl. Acad. Sci. USA* (1987) 84:7413.

Mammalian cell lines available as hosts for 15 expression are also known and include many immortalized cell lines available from the American Type Culture Collection (ATCC), including but not limited to, Chinese hamster ovary (CHO) cells, HeLa cells, baby hamster kidney (BHK) cells, monkey kidney cells (COS), human 20 hepatocellular carcinoma cells (e.g., Hep G2), human embryonic kidney cells, baby hamster kidney cells, mouse sertoli cells, canine kidney cells, buffalo rat liver cells, human lung cells, human liver cells, mouse mammary tumor cells, as well as others. The mammalian host 25 cells used to produce the target polypeptide of this invention may be cultured in a variety of media. Commercially available media such as Ham's F10 (Sigma), Minimal Essential Medium ([MEM], Sigma), RPMI-1640 (Sigma), and Dulbecco's Modified Eagle's Medium ([DMEM], Sigma) are suitable for culturing the host cells. In 30 addition, any of the media described in Ham and Wallace, *Meth. Enz.* (1979) 58: 44, Barnes and Sato, *Anal. Biochem.* (1980) 102: 255, U.S. Patent Nos. 4,767,704, 4,657,866, 4,927,762, or 4,560,655, WO 90/103430, WO 87/00195, and 35 U.S. RE 30,985, may be used as culture media for the host cells. Any of these media may be supplemented as necessary with hormones and/or other growth factors such

as insulin, transferrin, or epidermal growth factor, salts (such as sodium chloride, calcium, magnesium, and phosphate), buffers (such as HEPES), nucleosides (such as adenosine and thymidine), antibiotics (such as 5 Gentamycin(tm) M drug), trace elements (defined as inorganic compounds usually present at final concentrations in the micromolar range), and glucose or an equivalent energy source. Any other necessary supplements may also be included at appropriate 10 concentrations that would be known to those skilled in the art. The culture conditions, such as temperature, pH, and the like, are those previously used with the host cell selected for expression, and will be apparent to the ordinarily skilled artisan.

15 The active Factor VIII:C analogs produced according to the invention have a variety of uses. For example, the analogs can be used as immunogens for the production of antibodies. The analogs can also be used for the treatment of hemophiliacs and other hosts having 20 blood clotting disorders. In this regard, due to their resistance to proteolytic cleavage, the Factor VIII:C analogs may display increased plasma half-life or specific activity. Thus, the analogs may allow for lower dosages or alternative modes of administration and may 25 improve hemostasis in hemophiliacs.

Alternatively, nucleic acid molecules or vectors comprising polynucleotide sequences encoding the Factor VIII:C analogs can be used directly for gene therapy and administered using standard gene delivery 30 protocols. In this regard, the nucleotide sequences encoding the Factor VIII:C analogs can be stably integrated into the host cell genome or maintained on a stable episomal element in the host cell. Methods for gene delivery are known in the art. See, e.g., U.S. 35 Patent No. 5,399,346.

A number of viral based systems have been developed for gene transfer into mammalian cells. For

example, retroviruses provide a convenient platform for gene delivery systems. A selected gene can be inserted into a vector and packaged in retroviral particles using techniques known in the art. The recombinant virus can 5 then be isolated and delivered to cells of the subject either *in vivo* or *ex vivo*. A number of retroviral systems have been described (U.S. Patent No. 5,219,740; Miller and Rosman, *BioTechniques* (1989) 7:980-990; Miller, A.D., *Human Gene Therapy* (1990) 1:5-14; Scarpa et 10 al., *Virology* (1991) 180:849-852; Burns et al., *Proc. Natl. Acad. Sci. USA* (1993) 90:8033-8037; and Boris-Lawrie and Temin, *Cur. Opin. Genet. Develop.* (1993) 3:102-109. A number of adenovirus vectors have also been described. Unlike retroviruses which integrate into the 15 host genome, adenoviruses persist extrachromosomally thus minimizing the risks associated with insertional mutagenesis (Haj-Ahmad and Graham, *J. Virol.* (1986) 57:267-274; Bett et al., *J. Virol.* (1993) 67:5911-5921; Mittereder et al., *Human Gene Therapy* (1994) 5:717-729; 20 Seth et al., *J. Virol.* (1994) 68:933-940; Barr et al., *Gene Therapy* (1994) 1:51-58; Berkner, K.L. *BioTechniques* (1988) 6:616-629; and Rich et al., *Human Gene Therapy* (1993) 4:461-476).

Additionally, various adeno-associated virus 25 (AAV) vector systems have been developed for gene delivery. Such systems can include control sequences, such as promoter and polyadenylation sites, as well as selectable markers or reporter genes, enhancer sequences, and other control elements which allow for the induction 30 of transcription. AAV vectors can be readily constructed using techniques well known in the art. See, e.g., U.S. Patent Nos. 5,173,414 and 5,139,941; International Publication Nos. WO 92/01070 (published 23 January 1992) and WO 93/03769 (published 4 March 1993); Lebkowski et 35 al., *Molec. Cell. Biol.* (1988) 8:3988-3996; Vincent et al., *Vaccines* 90 (1990) (Cold Spring Harbor Laboratory Press); Carter, B.J. *Current Opinion in Biotechnology*

(1992) 3:533-539; Muzychka, N. *Current Topics in Microbiol. and Immunol.* (1992) 158:97-129; Kotin, R.M. *Human Gene Therapy* (1994) 5:793-801; Shelling and Smith, *Gene Therapy* (1994) 1:165-169; and Zhou et al., *J. Exp. 5 Med.* (1994) 179:1867-1875.

Additional viral vectors which will find use for delivering the nucleic acid molecules encoding the Factor VIII:C analog polypeptides for gene transfer include those derived from the pox family of viruses, 10 including vaccinia virus and avian poxvirus. By way of example, vaccinia virus recombinants expressing the novel Factor VIII:C analogs can be constructed as follows. The DNA encoding the particular analog is first inserted into an appropriate vector so that it is adjacent to a 15 vaccinia promoter and flanking vaccinia DNA sequences, such as the sequence encoding thymidine kinase (TK). This vector is then used to transfect cells which are simultaneously infected with vaccinia. Homologous recombination serves to insert the vaccinia promoter plus 20 the gene encoding the instant protein into the viral genome. The resulting TK⁻recombinant can be selected by culturing the cells in the presence of 5- bromodeoxyuridine and picking viral plaques resistant thereto.

25 A vaccinia based infection/transfection system can be conveniently used to provide for inducible, transient expression of the Factor VIII:C analogs in a host cell. In this system, cells are first infected *in vitro* with a vaccinia virus recombinant that encodes the 30 bacteriophage T7 RNA polymerase. This polymerase displays exquisite specificity in that it only transcribes templates bearing T7 promoters. Following infection, cells are transfected with the polynucleotide of interest, driven by a T7 promoter. The polymerase 35 expressed in the cytoplasm from the vaccinia virus recombinant transcribes the transfected DNA into RNA which is then translated into protein by the host

translational machinery. The method provides for high level, transient, cytoplasmic production of large quantities of RNA and its translation products. See, e.g., Elroy-Stein and Moss, *Proc. Natl. Acad. Sci. USA* 5 (1990) 87:6743-6747; Fuerst et al., *Proc. Natl. Acad. Sci. USA* (1986) 83:8122-8126.

Alternatively, avipoxviruses, such as the fowlpox and canarypox viruses, can also be used to deliver the Factor VIII:C analog genes. Recombinant 10 avipox viruses, expressing immunogens from mammalian pathogens, are known to confer protective immunity when administered to non-avian species. The use of an avipox vector is particularly desirable in human and other mammalian species since members of the avipox genus can 15 only productively replicate in susceptible avian species and therefore are not infective in mammalian cells. Methods for producing recombinant avipoxviruses are known in the art and employ genetic recombination, as described above with respect to the production of vaccinia viruses. 20 See, e.g., WO 91/12882; WO 89/03429; and WO 92/03545.

Molecular conjugate vectors, such as the adenovirus chimeric vectors described in Michael et al., *J. Biol. Chem.* (1993) 268:6866-6869 and Wagner et al., *Proc. Natl. Acad. Sci. USA* (1992) 89:6099-6103, can also 25 be used for gene delivery.

As an alternative approach to infection with vaccinia or avipox virus recombinants, or to the delivery of genes using other viral vectors, an amplification system can be used that will lead to high level 30 expression following introduction into host cells.

Specifically, a T7 RNA polymerase promoter preceding the coding region for T7 RNA polymerase can be engineered. Translation of RNA derived from this template will generate T7 RNA polymerase which in turn will transcribe 35 more template. Concomitantly, there will be a cDNA whose expression is under the control of the T7 promoter. Thus, some of the T7 RNA polymerase generated from

translation of the amplification template RNA will lead to transcription of the desired gene. Because some T7 RNA polymerase is required to initiate the amplification, T7 RNA polymerase can be introduced into cells along with 5 the template(s) to prime the transcription reaction.

The amplification template can be generated by PCR techniques. However the use of a plasmid is preferred. Since high level expression of T7 RNA polymerase appears to be lethal to host cells, the 10 plasmid should be one where expression of T7 RNA polymerase can be controlled. For example, a lac operator can be engineered distal or proximal (or both) to the T7 promoter. The binding of the preexisting lac repressor in the appropriate bacterial strain would 15 interfere with the transcription of the template by blocking access to the promoter by T7 RNA polymerase. Alternatively, or in combination with the above, a plasmid can be constructed where transcription from a bacterial promoter begins 3' of the T7 gene and continues 20 through the 5' end of the T7 promoter. Such transcription will generate an antisense transcript and reduce or eliminate translation of T7 RNA polymerase RNAs. The second transcription unit consisting of the T7 promoter preceding the gene of interest can be provided 25 by a separate plasmid or can be engineered onto the amplification plasmid. Colocalization of the two transcription units is beneficial for ease of manufacturing and ensures that both transcription units will always be together in the cells into which the 30 plasmid is introduced. The T7 RNA polymerase plasmids may include UTRs which comprise an Internal Ribosome Entry Site (IRES) present in the leader sequences of picornaviruses such as the encephalomyocarditis virus (EMCV) UTR (Jang et al. J. Virol. (1989) 63:1651-1660). 35 This sequence serves to enhance expression of sequences under the control of the T7 promoter. For a further discussion of T7 systems and their use for transforming

cells, see, e.g., International Publication No. WO 94/26911; Studier and Moffatt, *J. Mol. Biol.* (1986) 189:113-130; Deng and Wolff, *Gene* (1994) 143:245-249; Gao et al., *Biochem. Biophys. Res. Commun.* (1994) 200:1201-5 1206; Gao and Huang, *Nuc. Acids Res.* (1993) 21:2867-2872; Chen et al., *Nuc. Acids Res.* (1994) 22:2114-2120; and U.S. Patent No. 5,135,855.

Vectors encoding the subject Factor VIII:C analogs can also be packaged in liposomes prior to 10 delivery to the subject or to cells derived therefrom. Lipid encapsulation is generally accomplished using liposomes which are able to stably bind or entrap and retain nucleic acid. The ratio of condensed DNA to lipid preparation can vary but will generally be around 1:1 (mg 15 DNA:micromoles lipid), or more of lipid. For a review of the use of liposomes as carriers for delivery of nucleic acids, see, Hug and Sleight, *Biochim. Biophys. Acta.* (1991) 1097:1-17; Straubinger et al., in *METHODS OF ENZYMOLOGY* (1983), Vol. 101, pp. 512-527.

20 Liposomal preparations for use in the instant invention include cationic (positively charged), anionic (negatively charged) and neutral preparations, with cationic liposomes particularly preferred. Cationic liposomes have been shown to mediate intracellular 25 delivery of plasmid DNA (Felgner et al., *Proc. Natl. Acad. Sci. USA* (1987) 84:7413-7416); mRNA (Malone et al., *Proc. Natl. Acad. Sci. USA* (1989) 86:6077-6081); and purified transcription factors (Debs et al., *J. Biol. Chem.* (1990) 265:10189-10192), in functional form.

30 Cationic liposomes are readily available. For example, N[1-2,3-dioleyloxy)propyl]-N,N,N-triethyl-ammonium (DOTMA) liposomes are available under the trademark Lipofectin, from GIBCO BRL, Grand Island, NY. (See, also, Felgner et al., *Proc. Natl. Acad. Sci. USA* 35 (1987) 84:7413-7416). Other commercially available liposomes include transfectace (DDAB/DOPE) and DOTAP/DOPE (Boerhinger). Other cationic liposomes can be prepared

from readily available materials using techniques well known in the art. See, e.g., Szoka et al., *Proc. Natl. Acad. Sci. USA* (1978) 75:4194-4198; PCT Publication No. WO 90/11092 for a description of the synthesis of 5 DOTAP (1,2-bis(oleoyloxy)-3-(trimethylammonio)propane) liposomes.

Similarly, anionic and neutral liposomes are readily available, such as from Avanti Polar Lipids (Birmingham, AL), or can be easily prepared using readily 10 available materials. Such materials include phosphatidyl choline, cholesterol, phosphatidyl ethanolamine, dioleoylphosphatidyl choline (DOPC), dioleoylphosphatidyl glycerol (DOPG), dioleoylphosphatidyl ethanolamine (DOPE), among others. These materials can also be mixed with the 15 DOTMA and DOTAP starting materials in appropriate ratios. Methods for making liposomes using these materials are well known in the art.

The liposomes can comprise multilammelar vesicles (MLVs), small unilamellar vesicles (SUVs), or 20 large unilamellar vesicles (LUVs). The various liposome-nucleic acid complexes are prepared using methods known in the art. See, e.g., Straubinger et al., in *METHODS OF IMMUNOLOGY* (1983), Vol. 101, pp. 512-527; Szoka et al., *Proc. Natl. Acad. Sci. USA* (1978) 75:4194-4198; 25 Papahadjopoulos et al., *Biochim. Biophys. Acta* (1975) 394:483; Wilson et al., *Cell* (1979) 17:77; Deamer and Bangham, *Biochim. Biophys. Acta* (1976) 443:629; Ostro et al., *Biochem. Biophys. Res. Commun.* (1977) 76:836; Fraley et al., *Proc. Natl. Acad. Sci. USA* (1979) 76:3348; Enoch 30 and Strittmatter, *Proc. Natl. Acad. Sci. USA* (1979) 76:145; Fraley et al., *J. Biol. Chem.* (1980) 255:10431; Szoka and Papahadjopoulos, *Proc. Natl. Acad. Sci. USA* (1978) 75:145; and Schaefer-Ridder et al., *Science* (1982) 215:166.

35 The recombinant vectors (whether or not encapsulated in liposomes), may be administered in pharmaceutical compositions as described above. The

pharmaceutical compositions will comprise sufficient genetic material to produce a therapeutically effective amount of the analog or analogs, as described above. For purposes of the present invention, an effective dose will 5 be from about 0.05 mg/kg to about 50 mg/kg of the DNA constructs in the individual to which it is administered. Once formulated, the compositions of the invention can be administered directly to the subject or, alternatively, in the case of the vectors described above, delivered ex 10 vivo, to cells derived from the subject. Methods for the ex vivo delivery and reimplantation of transformed cells into a subject are known in the art and described in e.g., International Publication No. WO 93/14778 (published 5 August 1993). Generally, such methods will 15 include dextran-mediated transfection, calcium phosphate precipitation, polybrene mediated transfection, protoplast fusion, electroporation, encapsulation of the polynucleotide(s) in liposomes, and direct microinjection of the DNA into nuclei, all well known in the art.

20 Direct delivery of the compositions will generally be accomplished by injection, either subcutaneously, intraperitoneally, intravenously or intramuscularly. Other modes of administration include oral and pulmonary administration, suppositories, and 25 transdermal applications. Dosage treatment may be a single dose schedule or a multiple dose schedule.

30 Although any similar or equivalent methods and materials may be employed in the practice or testing of the present invention, the preferred methods and materials are now described.

The present invention will now be illustrated by reference to the following examples which set forth particularly advantageous embodiments. However, it should be noted that these embodiments are illustrative 35 and are not to be construed as restricting the invention in any way.

Examples

The Factor VIII:C polypeptide analogs are made using conventional mutagenesis techniques. In this regard, mutagenesis of the Factor VIII:C nucleotide sequences can be performed utilizing plasmids which include sequences encoding the full-length molecule, plasmids encoding the light and heavy chains and various modifications of these molecules, depending on the Factor VIII:C analog desired. Such plasmids are known and described in, e.g., U.S. Patent no. 5,045,455. The plasmids are first linearized e.g., by using restriction endonucleases which cleave at unique restriction sites. Once linearized, the plasmids are treated with calf intestine phosphatase and separated on low melting temperature tris-acetate agarose gels. The linearized band is extracted by adsorption to silica dioxide and eluted in tris-EDTA. The plasmid is then denatured and the desired phosphorylated mutagenic oligonucleotide is added. The mixture is heated and allowed to slowly cool at room temperature. A heteroduplex oligonucleotide mixture can be used and the reactions made with, e.g., 2 mM MgCl₂, 1mM beta-mercaptoethanol, 400 μ M ATP, 100 μ M deoxynucleotide triphosphate, 3-4 units/ μ L of Klenow fragment of *E. coli* DNA polymerase I and 400 units/ μ L of T4 DNA ligase.

The reactions are terminated using phenolchloroform extraction and ethanol precipitation. DNA obtained is used to transform bacterial host cells and positive clones selected. DNA from the clones is transferred to nitrocellulose, filters prepared, and hybridized to screening probes to ensure that the mutagenic oligonucleotide is introduced into the correct fragment. Final mutations are confirmed by DNA sequencing.

The DNA can be prepared by banding in CsCl and can be used to transfect COS-1 monkey cells as described in Kaufman, PNAS (1982) 82:689. After transfection, the

polypeptide analog is isolated and Factor VIII:C activity is assayed by the Kabi Coatest chromogenic assay method for the ability to clot Factor VIII deficient plasma before and after thrombin activation.

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Example 1

Construction of a Factor VIII:C Polypeptide Analog Pro221

Residue 221 of Factor VIII:C is mutated using the oligonucleotide TTC ATG CAG GAT AGG CCX GCT GCA TCT 10 GCT CGG. The GAT encoding for Asp which normally occurs at the underlined position is mutated to form the codon for Pro which can be CCX where X is A, T, G or C. The mutagenesis is carried out, the correct sequence is confirmed, and the analog produced and tested for 15 activity, as described above.

Other Factor VIII:C polypeptide analogs can be constructed and assayed as described above.

The present invention has been described with reference to specific embodiments. However, this 20 application is intended to cover those changes and substitutions which may be made by those skilled in the art without departing from the spirit and the scope of the appended claims.

WHAT IS CLAIMED IS:

1. An active Factor VIII:C polypeptide analog comprising a native Factor VIII:C polypeptide that is 5 modified at a site adjacent to a non-activating Arg residue, wherein the modification comprises creation of an Arg-Pro or a Pro-Arg dipeptide.

2. The analog of claim 1, wherein the non- 10 activating Arg residue is an Arg residue selected from the group consisting of amino acid residues 220, 226, 250, 279, 282, 336, 359, 562, 747, 776, 1310, 1313, 1645, 1648, 1719, and 1721, numbered with respect to the native Factor VIII:C polypeptide sequence.

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3. The analog of claim 1, wherein the modification is an amino acid addition, substitution, or deletion, or a combination thereof.

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4. The analog of claim 1, wherein the modification is amino acid substitution and the substitution is of about one to about 7 amino acid residues.

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5. The analog of claim 1, wherein the modification is amino acid deletion, and the deletion is of about one to about 7 amino acid residues.

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6. An active Factor VIII:C polypeptide analog comprising a native Factor VIII:C polypeptide that is modified at a site adjacent to a non-activating Arg residue, wherein the modification comprises creation of a tripeptide having the formula $P_3-P_2-P_1$, wherein P_3 is a residue selected from the group consisting of Phe, Glu, 35 and Pro; P_2 is any amino acid residue except Ser and P_2 is not Leu335 and is not Asn1720; and P_1 is Arg.

7. The analog of claim 2, wherein the non-activating Arg residue is Arg220, and the modification comprises at least one selected from the group consisting of: (a) an insertion of at least one Pro residue between 5 Asp219 and Arg 220; (b) an insertion of at least one Pro residue between Arg220 and Asp221; and (c) a deletion comprising residues Asp221, Ala222, Ala223, Ser224, Ala225, Arg226, Ala227, and Trp228.

10 8. The analog of claim 6, wherein the non-activating Arg residue is Arg220, and the modification comprises at least one selected from the group consisting of: (a) an insertion of at least one Phe residue between Gln218 and Asp219; (b) an insertion of at least one Glu residue between Gln218 and Asp219; (c) an insertion of at 15 least one Pro residue between Gln218 and Asp219; (d) a deletion comprising residues Thr212, Lys213, Asn214, Ser215, Leu216, Met217, and Gln218; (e) a substitution of at least one Phe residue at Gln218; (f) a substitution of 20 at least one Glu residue at Gln218; and (g) a substitution of at least one Pro residue at Gln218.

9. The analog of claim 2, wherein the non-activating Arg residue is Arg226, and the modification 25 comprises at least one selected from the group consisting of: (a) an insertion of at least one Pro residue between Ala225 and Arg226; (b) an insertion of at least one Pro residue between Arg226 and Ala227; (c) a deletion comprising residues Ala227 and Trp228; (d) a deletion 30 comprising residues Ala227 and Trp228 and insertion of at least one Pro residue to replace the deleted residues; and (e) a deletion comprising residues Ala227, Trp228, Pro229, and Lys230, and an insertion of at least one Pro residue to replace the deleted residues.

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10. The analog of claim 6, wherein the non-activating Arg residue is Arg226, and the modification

comprises at least one selected from the group consisting of: (a) an insertion of at least one Phe residue between Ser224 and Ala225; (b) an insertion of at least one Glu residue between Ser224 and Ala225; (c) an insertion of at 5 least one Pro residue between Ser224 and Ala225; (d) a substitution of at least one Phe residue at Ser224; (e) a substitution of at least one Glu residue at Ser224; and (f) a substitution of at least one Pro residue at Ser224.

10 11. The analog of claim 2, wherein the non-activating Arg residue is Arg250, and the modification comprises at least one selected from the group consisting of: (a) an insertion of at least one Pro residue between His249 and Arg250; (b) an insertion of at least one Pro 15 residue between Arg250 and Lys251; (c) a deletion comprising residues Lys251 and Ser252 and insertion of at least one Pro residue to replace the deleted residues; (d) a substitution of Lys251 with at least one Pro residue; (e) a deletion comprising residues Gly244, 20 Leu245, Ile246, Gly247, Cys248, and His249; and (f) a deletion comprising residues Arg240, Ser241, Leu242, Pro243, Gly244, Leu245, Ile246, Gly247, Cys248, and His249 and an insertion of at least one Pro residue to replace the deleted residues.

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12. The analog of claim 6, wherein the non-activating Arg residue is Arg250, and the modification comprises at least one selected from the group consisting of: (a) an insertion of at least one Phe residue between 30 Cys248 and His249; (b) an insertion of at least one Glu residue between Cys248 and His249; (c) an insertion of at least one Pro residue between Cys248 and His249; (d) a deletion comprising residues Gly244, Leu245, Ile246, Gly247, and Cys248; (e) a substitution of at least one 35 Phe residue at Cys248; (f) a substitution of at least one Glu residue at Cys248; and (g) a substitution of at least one Pro residue at Cys248.

13. The analog of claim 2, wherein the non-activating Arg residue is Arg279, and the modification comprises at least one selected from the group consisting of: (a) an insertion of at least one Pro residue between 5 Val278 and Arg279; (b) an insertion of at least one Pro residue between Arg279 and Asn280; (c) a deletion comprising residues Asn280, His281, and Arg282 and an insertion of at least one Pro residue to replace the deleted residues; and (d) a deletion comprising Asn280, 10 His281, Arg282, Gln283, Ala284, and Ser285, and an insertion of at least one Pro residue to replace the deleted residues.

14. The analog of claim 6, wherein the non-activating Arg residue is Arg279, and the modification comprises at least one selected from the group consisting of: (a) an insertion of at least one Phe residue between 15 Leu277 and Val278; (b) an insertion of at least one Glu residue between Leu277 and Val278; (c) an insertion of at least one Pro residue between Leu277 and Val278; (d) a deletion comprising residue Leu277; (e) a deletion comprising residue Val278; (f) a deletion comprising residues Gly273, His274, Thr275, Phe276, and Leu277; (g) a deletion comprising residues Leu271, Glu272, Gly273, 20 His274, Thr275, Phe276, and Leu277; (h) a substitution of at least one Phe residue at Leu277; (i) a substitution of at least one Glu residue at Leu277; and (j) a 25 substitution of at least one Pro residue at Leu277.

30 15. The analog of claim 2, wherein the non-activating Arg residue is Arg282, and the modification comprises at least one selected from the group consisting of: (a) an insertion of at least one Pro residue between His281 and Arg282; (b) an insertion of at least one Pro 35 residue between Arg282 and Gln283; (c) a deletion comprising residues Arg279, Asn280, and His281 and an insertion of at least one Pro residue to replace the

deleted residues; (d) a deletion comprising residues Gln283, Ala284, and Ser285, and an insertion of at least one Pro residue to replace the deleted residues; and (e) a deletion comprising residues Gln283, Ala284, Ser285, 5 Leu286, Glu287, Ile288, and Ser289.

16. The analog of claim 6, wherein the non-activating Arg residue is Arg282, and the modification comprises at least one selected from the group consisting 10 of: (a) an insertion of at least one Phe residue between Asn280 and His281; (b) an insertion of at least one Glu residue between Asn280 and His281; (c) an insertion of at least one Pro residue between Asn280 and His281; (d) a deletion comprising residues Leu277, Val278, Arg279, and 15 Asn280; (e) a deletion comprising residues Gly273, His274, Thr275, Phe276, Leu277, Val278, Arg279, and Asn280; (f) a substitution of at least one Phe residue at Asn280; (g) a substitution of at least one Glu residue at Asn280; and (h) a substitution of at least one Pro 20 residue at Asn280.

17. The analog of claim 2, wherein the non-activating Arg residue is Arg336, and the modification comprises at least one selected from the group consisting 25 of: (a) an insertion of at least one Pro residue between Leu335 and Arg336; (b) a deletion comprising residues Gln334, and Leu335; (c) an insertion of at least one Pro residue between Arg336 and Met337; and (d) a deletion comprising residues Met337, and Lys338 and insertion of 30 at least one Pro residue in place of the deleted residues.

18. The analog of claim 6, wherein the non-activating Arg residue is Arg336, and the modification 35 comprises at least one selected from the group consisting of: (a) a deletion comprising residue Leu335 and an insertion of at least one Phe residue between Pro333 and

Gln334; (b) a deletion comprising residue Leu335 and an insertion of at least one Glu residue between Pro333 and Gln334; (c) a deletion comprising residue Leu335 and an insertion of at least one Pro residue between Pro333 and 5 Gln334; (d) a deletion comprising residue Leu335; (e) a deletion comprising residues Gln334, and Leu335; (f) a deletion comprising residues Pro333, Gln334, and Leu335; (g) a deletion comprising residues Glu332, Pro333, Gln334, and Leu335; (h) a deletion comprising residue 10 Leu335 and a substitution of at least one Phe residue at Pro333; and (i) a deletion comprising residue Leu335 and a substitution of at least one Glu residue at Pro333.

19. The analog of claim 2, wherein the non-activating Arg residue is Arg359, and the modification comprises at least one selected from the group consisting of: (a) an insertion of at least one Pro residue between Val358 and Arg359; (b) an insertion of at least one Pro residue between Arg359 and Phe360; (c) a deletion 20 comprising residues Phe360, Asp361, Asp362, Asp363, Asn364, and Ser365; and (d) a deletion comprising residues Phe360, Asp361, Asp362, Asp363, Asn364, Ser365, Pro366, and Ser367, and an insertion of at least one Pro residue to replace the deleted residues.

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20. The analog of claim 6, wherein the non-activating Arg residue is Arg359, and the modification comprises at least one selected from the group consisting of: (a) an insertion of at least one Phe residue between Val357 and Val358; (b) an insertion of at least one Glu residue between Val357 and Val358; (c) an insertion of at least one Pro residue between Val357 and Val358; (d) a deletion comprising residues Met355, Asp356, and Val357; (e) a substitution of at least one Phe residue at Val357; 30 (f) a substitution of at least one Glu residue at Val357; and (g) a substitution of at least one Pro residue at Val357; 35 and (h) a substitution of at least one Glu residue at Val357.

21. The analog of claim 2, wherein the non-activating Arg residue is Arg562, and the modification comprises at least one selected from the group consisting of: (a) an insertion of at least one Pro residue between 5 Gln561 and Arg562; (b) an insertion of at least one Pro residue between Arg562 and Gly563; (c) a deletion comprising residues Ser558, Val559, Asp560, Gln561, and an insertion of at least one Pro residue to replace the deleted residues; (d) a deletion comprising residues 10 Lys556, Glu557, Ser558, Val559, Asp560, Gln561, and an insertion of at least one Pro residue to replace the deleted residues; (e) a deletion comprising residues Gly563, Asn564, Gln565, Ile566, Met567, and Ser568, and an insertion of at least one Pro residue to replace the 15 deleted residues; and (f) a deletion comprising residues Gly563, Asn564, Gln565, Ile566, Met567, Ser568, Asp569, Lys570, and Arg571, and an insertion of at least one Pro residue to replace the deleted residues.

20 22. The analog of claim 6, wherein the non-activating Arg residue is Arg562, and the modification comprises at least one selected from the group consisting of: (a) an insertion of at least one Phe residue between Asp560 and Gln561; (b) an insertion of at least one Glu residue between Asp560 and Gln561; (c) an insertion of at least one Pro residue between Asp560 and Gln561; (d) a deletion comprising residues Ser558, Val559, and Asp560; (e) a substitution of at least one Phe residue at Asp560; (f) a substitution of at least one Glu residue at Asp560; 30 and (g) a substitution of at least one Pro residue at Asp560.

23. The analog of claim 2, wherein the non-activating Arg residue is Arg747, and the modification 35 comprises at least one selected from the group consisting of: (a) an insertion of at least one Pro residue between Ser746 and Arg747; (b) a deletion comprising residue

Ser746 and an insertion of at least one Pro residue to replace the deleted residue; (c) a deletion comprising residue His748; (d) a deletion comprising residue His748 and an insertion of at least one Pro residue to replace 5 the deleted residue; (e) an insertion of at least one Pro residue between Arg747 and His748; (f) a deletion comprising residues His748, Pro749, and Ser750 and an insertion of at least one Pro residue to replace the deleted residues; (g) a deletion comprising Ser743, 10 Gln744, Asn745, Ser746; and (h) a deletion comprising His748, Pro749, Ser750, Thr751, Arg752, Gln753, and Lys754, and an insertion of at least one Pro residue to replace the deleted residues.

15 24. The analog of claim 6, wherein the non-activating Arg residue is Arg747, and the modification comprises at least one selected from the group consisting of: (a) a deletion comprising residue Ser746 and an insertion of at least one Phe residue between Gln744 and 20 Asn745; (b) a deletion comprising residue Ser746 and an insertion of at least one Glu residue between Gln744 and Asn745; (c) a deletion comprising residue Ser746 and an insertion of at least one Pro residue between Gln744 and Asn745; (d) a deletion comprising residue Ser746 and a 25 substitution of at least one Phe residue at Gln744; (e) a deletion comprising residue Ser746 and a substitution of at least one Glu residue at Gln744; and (f) a deletion comprising residue Ser746 and a substitution of at least one Pro residue at Gln744.

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25. The analog of claim 2, wherein the non-activating Arg residue is Arg776, and the modification comprises at least one selected from the group consisting of: (a) an insertion of at least one Pro residue between 35 His775 and Arg776; (b) an insertion of at least one Pro residue between Arg776 and Thr777; (c) a deletion comprising residue Thr777; (d) a deletion comprising

residue Thr777 and an insertion of at least one Pro residue to replace the deleted residue; (e) a deletion comprising residues Trp772, Phe773, Ala774, and His775; (f) a deletion comprising residues Trp772, Phe773, 5 Ala774, and His775, and an insertion of at least one Pro residue to replace the deleted residues; (g) a deletion comprising residues Lys768, Thr769, Asp770, Pro771, Trp772, Phe773, Ala774, and His775, and an insertion of at least one Pro residue to replace the deleted residues; 10 and (h) a deletion comprising residues Thr777, Pro778, met779, Pro780, and Lys781, and an insertion of at least one Pro residue to replace the deleted residues.

26. The analog of claim 6, wherein the non-15 activating Arg residue is Arg776, and the modification comprises at least one selected from the group consisting of: (a) an insertion of at least one Phe residue between Ala774 and His775; (b) an insertion of at least one Glu residue between Ala774 and His775; (c) an insertion of at 20 least one Pro residue between Ala774 and His775; (d) a deletion comprising residue Ala774; (e) a deletion comprising residues Trp772, Phe773 and Ala774; (f) a deletion comprising residues Lys768, Thr769, Asp770, Pro771, Trp772, and Phe773; (g) a substitution of at 25 least one Phe residue at Ala774; (h) a substitution of at least one Glu residue at Ala774; and (i) a substitution of at least one Pro residue at Ala774.

27. The analog of claim 2, wherein the non-30 activating Arg residue is Arg1310, and the modification comprises at least one selected from the group consisting of: (a) an insertion of at least one Pro residue between Gln1309 and Arg1310; (b) an insertion of at least one Pro residue between Arg1310 and Ser1311; (c) a deletion 35 comprising residue Ser1311 and an insertion of at least one Pro residue to replace the residue; (d) a deletion comprising residues Ser1311, Lys1312, and Arg1313 and an

insertion of at least one Pro residue to replace the deleted residues; (e) a deletion comprising residues Ser1311, Lys1312, Arg1313, Ala1314, Leu1315, and Lys1316, and an insertion of at least one Pro residue to replace 5 the deleted residues; (f) a deletion comprising residues Ser1311, Lys1312, Arg1313, Ala1314, Leu1315, Lys1316, Gln1317, Phe1318, Arg1319, and Leu1320; and (g) a deletion comprising residues Ser1311, Lys1312, Arg1313, Ala1314, Leu1315, Lys1316, Gln1317, Phe1318, Arg1319, and 10 Leu1320, and an insertion of at least one Pro residue to replace the deleted residues.

28. The analog of claim 6, wherein the non-activating Arg residue is Arg1310, and the modification 15 comprises at least one selected from the group consisting of: (a) an insertion of at least one Phe residue between Thr1308 and Gln1309; (b) an insertion of at least one Glu residue between Thr1308 and Gln1309; (c) an insertion of at least one Pro residue between Thr1308 and Gln1309; (d) 20 a deletion comprising residues Val1307, and Thr1308; (e) a substitution of at least one Phe residue at Thr1308; (f) a substitution of at least one Glu residue at Thr1308; and (g) a substitution of at least one Pro residue at Thr1308.

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29. The analog of claim 2, wherein the non-activating Arg residue is Arg1313, and the modification comprises at least one selected from the group consisting of: (a) an insertion of at least one Pro residue between 30 Lys1312 and Arg1313; (b) a deletion comprising residue Lys1312 and an insertion of at least one Pro residue to replace the deleted residue; (c) a deletion comprising residues Arg1310, Ser1311, and Lys1312, and an insertion of at least one Pro residue to replace the deleted 35 residues; (d) an insertion of at least one Pro residue between Arg1313 and Ala1314; (e) a deletion comprising residues Ala1314, Leu1315, and Lys1316, and an insertion

of at least one Pro residue to replace the deleted residues; (f) a deletion comprising residues Ala1314, Leu1315, Lys1316, Gln1317, Phe1318, and Arg1319 and an insertion of at least one Pro residue to replace the 5 deleted residues; and (g) a deletion comprising residues Ala1314, Leu1315, Lys1316, Gln1317, Phe1318, Arg1319, and Leu1320.

30. The analog of claim 6, wherein the non-
10 activating Arg residue is Arg1313, and the modification comprises at least one selected from the group consisting of: (a) an insertion of at least one Phe residue between Ser1311 and Lys1312; (b) an insertion of at least one Glu residue between Ser1311 and Lys1312; (c) an insertion of 15 at least one Pro residue between Ser1311 and Lys1312; (d) a deletion comprising residues Val1307, Thr1308, Gln1309, Arg1310, and Ser1311; (e) a substitution of at least one Phe residue at Ser1311; (f) a substitution of at least one Glu residue at Ser 1311; and (g) a substitution of at 20 least one Pro residue at Ser 1311.

31. The analog of claim 2, wherein the non-activating Arg residue is Arg1645, and the modification comprises at least one selected from the group consisting 25 of: (a) a deletion comprising residues Val1642, Leu1643, and Lys1644; (b) a deleteion comprising residue Lys1644 and an insertion of at least one Pro residue to replace the deleted residue; (c) an insertion of at least one Pro residue between Arg1645 and Lys1644; (d) an insertion of 30 at least one Pro residue between Arg1645 and His1646; (e) a deletion comprising residues His1646, Gln1647, and Arg1648 and an insertion of at least one Pro residue to replace the deleted residues; and (f) a deletion comprising residues His1646, Gln1647, Arg1648, Glu1649, 35 Ile1650, Thr1651, and Arg1652, and an insertion of at least one Pro residue to replace the deleted residues.

32. The analog of claim 6, wherein the non-activating Arg residue is Arg1645, and the modification comprises at least one selected from the group consisting of: (a) an insertion of at least one Phe residue between 5 Leu1643 and Lys1644; (b) an insertion of at least one Glu residue between Leu1643 and Lys1644; (c) an insertion of at least one Pro residue between Leu1643 and Lys1644; (d) a deletion comprising residues Val1642, and Leu1643; (e) a deletion comprising residues Pro1641, Val1642, and 10 Leu1643; (f) a substitution of at least one Phe residue at Leu1643; (g) a substitution of at least one Glu residue at Leu1643; and (h) a substitution of at least one Pro residue at Leu1643.

15 33. The analog of claim 2, wherein the non-activating Arg residue is Arg1648, and the modification comprises at least one selected from the group consisting of: (a) a deletion comprising residues Val1642, Leu1643, Lys1644, Arg1645, His1646, and Gln1647; (b) an insertion 20 of at least one Pro residue between Gln1647 and Arg1648; (c) a deletion comprising residues Lys1644, Arg1645, His1646, and Gln1647 and an insertion of at least one Pro residue to replace the deleted residues; (d) a deletion comprising residues Glu1649, Ile1650, Thr1651, and 25 Arg1652 and an insertion of at least one Pro residue to replace the deleted residues; and (e) an insertion of at least one Pro residue between Arg1648 and Glu1649.

34. The analog of claim 6, wherein the non-activating Arg residue is Arg1648, and the modification comprises at least one selected from the group consisting of: (a) an insertion of at least one Phe residue between His1646 and Gln1647; (b) an insertion of at least one Glu residue between His1646 and Gln1647; (c) an insertion of 35 at least one Pro residue between His1646 and Gln1647; (d) a deletion comprising residues Val1642, Leu1643, Lys1644, Arg1645, and His1646; (e) a deletion comprising residues

Pro1641, Val1642, Leu1643, Lys1644, Arg1645, and His1646; (f) a substitution of at least one Phe residue at His1646; (g) a substitution of at least one Glu residue at His1646; and (h) a substitution of at least one Pro residue at His1646.

35. The analog of claim 2, wherein the non-activating Arg residue is Arg1719, and the modification comprises at least one selected from the group consisting of: (a) a deletion comprising residues His1716, Val1717, and Leu1718; (b) an insertion of at least one Pro residue between Leu1718 and Arg1719; (c) a deletion comprising residues Asn1720, and Arg1721 and an insertion of at least one Pro residue to replace the deleted residues; and (d) an insertion of at least one Pro residue between Arg1719 and Asn1720.

36. The analog of claim 6, wherein the non-activating Arg residue is Arg1719, and the modification comprises at least one selected from the group consisting of: (a) an insertion of at least one Phe residue between Val1717 and Leu1718; (b) an insertion of at least one Glu residue between Val1717 and Leu1718; (c) an insertion of at least one Pro residue between Val1717 and Leu1718; (d) a deletion comprising residues His1716, and Val1717; (e) a substitution of at least one Phe residue at Val1717; and (f) a substitution of at least one Glu residue at Val1717; (g) a substitution of at least one Pro residue at Val1717.

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37. The analog of claim 2, wherein the non-activating Arg residue is Arg1721, and the modification comprises at least one selected from the group consisting of: (a) a deletion comprising residues His1716, Val1717, Leu1718, Arg1719, and Asn1720; (b) an insertion of at least one Pro residue between Asn1720 and Arg1721; (c) an insertion of at least one Pro residue between Arg1721 and

Ala1722; (d) a deletion comprising residues Ala1722, Gln1723, and Ser1724 and an insertion of at least one Pro residue to replace the deleted residues; (e) a deletion comprising residues Ala1722, Gln1723, Ser1724, Gly1725, 5 Ser1726, and Val1727; and (f) a deletion comprising residues Ala1722, Gln1723, Ser1724, Gly1725, and Ser1726, and an insertion of at least one Pro residue to replace the deleted residues.

10 38. The analog of claim 6, wherein the non-activating Arg residue is Arg1721, and the modification comprises at least one selected from the group consisting of: (a) a deletion comprising residue Asn1720 and an insertion of at least one Phe residue between Leu1718 and 15 Arg1719; (b) a deletion comprising residue Asn1720 and an insertion of at least one Glu residue between Leu1718 and Arg1719; (c) a deletion comprising residue Asn1720 and an insertion of at least one Pro residue between Leu1718 and Arg1719; (d) a deletion comprising residues Val1717, 20 Leu1718, Arg1719, and Asn1720; (e) a deletion comprising residue Asn1720 and a substitution of at least one Phe residue at Leu1718; (f) a deletion comprising residue Asn1720 and a substitution of at least one Glu residue at Leu1718; and (g) a deletion comprising residue Asn1720 25 and a substitution of at least one Pro residue at Leu1718;

39. An active Factor VIII:C polypeptide analog comprising a native Factor VIII:C polypeptide that is 30 modified at at least one non-activating Arg residue selected from the group consisting of Arg336, Arg1719 and Arg1721, wherein the modification comprises a substitution of any of amino acids Pro, Glu, Asp, Asn, Gln, Ser and Tyr for Arg336, a substitution of any of 35 amino acids Pro, Glu, Asp, Asn, Gln, Ser and Tyr for Arg1719 and/or a substitution of any of amino acids Glu,

Asp, Asn, Gln, Ser and Tyr for Arg1721, numbered with respect to the native Factor VIII:C polypeptide sequence.

40. The analog of claim 39, wherein the 5 modification comprises at least one amino acid substitution selected from the group consisting of a substitution of Pro for Arg336, a substitution of Pro for Arg1719 and a substitution of Glu for Arg1721, numbered with respect to the native Factor VIII:C polypeptide 10 sequence.

41. The analog of claim 1, wherein the native Factor VIII:C polypeptide is selected from the group consisting of:

- 15 a) a full-length Factor VIII:C molecule comprising a signal peptide and all A, B, and C domains;
- b) a mature Factor VIII:C molecule comprising all A, B, and C domains and lacking a signal peptide;
- 20 c) a truncated Factor VIII:C molecule lacking a signal peptide and at least a portion of the B domain;
- d) a cleaved Factor VIII:C molecule comprising a light chain subunit of molecular weight of 25 about 80 kD;
- e) a cleaved Factor VIII:C molecule comprising a heavy chain fragment of molecular weight in a range of about 90 kD to 200 kD;
- 30 f) a cleaved Factor VIII:C molecule comprising a heavy chain fragment of molecular weight of about 90 kD;
- g) a cleaved Factor VIII:C molecule comprising a heavy chain fragment of molecular weight of about 50 kD;
- 35 h) a cleaved Factor VIII:C molecule comprising a heavy chain fragment of molecular weight of about 43 kD; and

i) a cleaved Factor VIII:C molecule comprising a light chain fragment of molecular weight of about 73 kD.

5 42. An active Factor VIII:C polypeptide analog complex comprising at least two Factor VIII:C polypeptide analogs as claimed in claim 41, or at least one Factor VIII:C polypeptide analog and at least one Factor VIII:C polypeptide, wherein the complex comprises a metal ion.

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43. The analog complex of claim 41, wherein the complex comprises two Factor VIII:C polypeptide analogs, and the Factor VIII:C polypeptides that are modified to form the two analogs are selected from the 15 group consisting of molecular weights of about (a) 80 kD and 90 kD; (b) 73 kD and 90 kD; (c) 80 kD and 50 kD; (d) 80 kD and 43 kD; (e) 73 kD and 50 kD; and (f) 73 kD and 43 kD.

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44. The analog complex of claim 42, wherein the metal ion is a divalent cation.

45. The analog complex of claim 44, wherein the divalent cation is a Ca^{++} ion or a Cu^{++} ion.

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46. The analog of claim 6, wherein the native Factor VIII:C polypeptide is selected from the group consisting of:

30 a) a full-length Factor VIII:C molecule comprising a signal peptide and all A, B, and C domains;
b) a mature Factor VIII:C molecule comprising all A, B, and C domains and lacking a signal peptide;
c) a truncated Factor VIII:C molecule lacking a signal peptide and at least a portion of the B domain;

15 d) a cleaved Factor VIII:C molecule comprising a light chain subunit of molecular weight of about 80 kD;

20 e) a cleaved Factor VIII:C molecule comprising a heavy chain fragment of molecular weight in a range of about 90 kD to 200 kD;

25 f) a cleaved Factor VIII:C molecule comprising a heavy chain fragment of molecular weight of about 90 kD;

30 g) a cleaved Factor VIII:C molecule comprising a heavy chain fragment of molecular weight of about 50 kD;

35 h) a cleaved Factor VIII:C molecule comprising a heavy chain fragment of molecular weight of about 43 kD; and

40 i) a cleaved Factor VIII:C molecule comprising a light chain fragment of molecular weight of about 73 kD.

45 47. An active Factor VIII:C polypeptide analog complex comprising at least two Factor VIII:C polypeptide analogs as claimed in claim 46, or at least one Factor VIII:C polypeptide analog and at least one Factor VIII:C polypeptide, wherein the complex comprises a metal ion.

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55 48. The analog complex of claim 46, wherein the complex comprises two Factor VIII:C polypeptide analogs, and the Factor VIII:C polypeptides that are modified to form the two analogs are selected from the 60 group consisting of molecular weights of about (a) 80 kD and 90 kD; (b) 73 kD and 90 kD; (c) 80 kD and 50 kD; (d) 80 kD and 43 kD; (e) 73 kD and 50 kD; and (f) 73 kD and 43 kD.

65 49. The analog complex of claim 47, wherein the metal ion is a divalent cation.

50. The analog complex of claim 49, wherein the divalent cation is a Ca^{++} ion or a Cu^{++} ion.

51. The analog of claim 40, wherein the native Factor VIII:C polypeptide is selected from the group consisting of:

- a) a full-length Factor VIII:C molecule comprising a signal peptide and all A, B, and C domains;
- b) a mature Factor VIII:C molecule comprising all A, B, and C domains and lacking a signal peptide;
- c) a truncated Factor VIII:C molecule lacking a signal peptide and at least a portion of the B domain;
- d) a cleaved Factor VIII:C molecule comprising a light chain subunit of molecular weight of about 80 kD;
- e) a cleaved Factor VIII:C molecule comprising a heavy chain fragment of molecular weight in a range of about 90 kD to 200 kD;
- f) a cleaved Factor VIII:C molecule comprising a heavy chain fragment of molecular weight of about 90 kD;
- g) a cleaved Factor VIII:C molecule comprising a heavy chain fragment of molecular weight of about 50 kD;
- h) a cleaved Factor VIII:C molecule comprising a heavy chain fragment of molecular weight of about 43 kD; and
- i) a cleaved Factor VIII:C molecule comprising a light chain fragment of molecular weight of about 73 kD.

52. An active Factor VIII:C polypeptide analog complex comprising at least two Factor VIII:C polypeptide analogs as claimed in claim 51, or at least one Factor

VIII:C polypeptide analog and at least one Factor VIII:C polypeptide, wherein the complex comprises a metal ion.

53. The analog complex of claim 51, wherein
5 the complex comprises two Factor VIII:C polypeptide
analog, and the Factor VIII:C polypeptides that are
modified to form the two analogs are selected from the
group consisting of molecular weights of about (a) 80 kD
and 90 kD; (b) 73 kD and 90 kD; (c) 80 kD and 50 kD; (d)
10 80 kD and 43 kD; (e) 73 kD and 50 kD; and (f) 73 kD and
43 kD.

54. The analog complex of claim 52, wherein
the metal ion is a divalent cation.

15

55. The analog complex of claim 54, wherein
the divalent cation is a Ca^{++} ion or a Cu^{++} ion.

56. A method of producing a Factor VIII:C
20 polypeptide analog, comprising:

a) providing a native Factor VIII:C
polypeptide that comprises an amino acid sequence; and
b) modifying an amino acid residue in
the amino acid sequence to produce the analog of claim 1.

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57. A method of producing a Factor VIII:C
polypeptide analog, comprising:

a) providing a native Factor VIII:C
polypeptide that comprises an amino acid sequence; and
30 b) modifying an amino acid residue in
the amino acid sequence to produce the analog of claim 6.

58. A method of producing a Factor VIII:C
polypeptide analog, comprising:

35 a) providing a native Factor VIII:C
polypeptide that comprises an amino acid sequence; and

b) modifying an amino acid residue in the amino acid sequence to produce the analog of claim 40.

5 59. A nucleic acid molecule comprising a nucleotide sequence that encodes the Factor VIII:C polypeptide analog of claim 1.

10 60. A nucleic acid molecule comprising a nucleotide sequence that encodes the Factor VIII:C polypeptide analog of claim 6.

15 61. A nucleic acid molecule comprising a nucleotide sequence that encodes the Factor VIII:C polypeptide analog of claim 40.

20 62. A recombinant vector comprising the nucleic acid molecule of claim 59 and a regulatory element, wherein the nucleic acid molecule is under regulatory control of the regulatory element.

25 63. A recombinant vector comprising the nucleic acid molecule of claim 60 and a regulatory element, wherein the nucleic acid molecule is under regulatory control of the regulatory element.

30 64. A recombinant vector comprising the nucleic acid molecule of claim 61 and a regulatory element, wherein the nucleic acid molecule is under regulatory control of the regulatory element.

65. A recombinant host cell comprising the nucleic acid molecule of claim 59.

35 66. A recombinant host cell comprising the nucleic acid molecule of claim 60.

67. A recombinant host cell comprising the nucleic acid molecule of claim 61.

68. A recombinant host cell comprising the 5 recombinant vector of claim 62.

69. A recombinant host cell comprising the recombinant vector of claim 63.

10 70. A recombinant host cell comprising the recombinant vector of claim 64.

71. A method of producing an active Factor VIII:C polypeptide analog comprising:

15 a) providing the recombinant host cell of claim 65; and
b) allowing the recombinant host cell to express the analog.

20 72. A method of producing an active Factor VIII:C polypeptide analog comprising:

a) providing the recombinant host cell of claim 66; and
b) allowing the recombinant host cell to 25 express the analog.

73. A method of producing an active Factor VIII:C polypeptide analog comprising:

30 a) providing the recombinant host cell of claim 67; and
b) allowing the recombinant host cell to express the analog.

74. A method of producing an active Factor 35 VIII:C polypeptide analog comprising:

a) providing the recombinant host cell of claim 68; and

10 b) allowing the recombinant host cell to express the analog.

15 75. A method of producing an active Factor VIII:C polypeptide analog comprising:

20 a) providing the recombinant host cell of claim 69; and

25 b) allowing the recombinant host cell to express the analog.

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35 76. A method of producing an active Factor VIII:C polypeptide analog comprising:

40 a) providing the recombinant host cell of claim 70; and

45

50 b) allowing the recombinant host cell to express the analog.

55

77. A method of producing a nucleic acid molecule that encodes a Factor VIII:C polypeptide analog comprising:

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65 a) providing a nucleic acid molecule that encodes a native Factor VIII:C polypeptide, wherein the nucleic acid molecule comprises a codon for each amino acid residue in the native Factor VIII:C 70 polypeptide; and

75 b) modifying at least one codon that is adjacent to a codon encoding a non-activating Arg residue, to produce the nucleic acid molecule of claim 59.

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85 78. The method of producing a nucleic acid molecule as claimed in claim 77, wherein the modifying is performed by site directed mutagenesis or by use of polymerase chain reaction techniques.

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79. A method of producing a nucleic acid molecule that encodes a Factor VIII:C polypeptide analog comprising:

5 a) providing a nucleic acid molecule that encodes a native Factor VIII:C polypeptide, wherein the nucleic acid molecule comprises a codon for each amino acid residue in the native Factor VIII:C polypeptide; and

10 b) modifying at least one codon that is adjacent to a codon encoding a non-activating Arg residue, to produce the nucleic acid molecule of claim 60.

15 80. The method of producing a nucleic acid molecule as claimed in claim 79, wherein the modifying is performed by site directed mutagenesis or by use of polymerase chain reaction techniques.

20 81. A method of producing a nucleic acid molecule that encodes a Factor VIII:C polypeptide analog comprising:

25 a) providing a nucleic acid molecule that encodes a native Factor VIII:C polypeptide, wherein the nucleic acid molecule comprises a codon for each amino acid residue in the native Factor VIII:C polypeptide; and

30 b) modifying at least one codon that is adjacent to a codon encoding a non-activating Arg residue, to produce the nucleic acid molecule of claim 61.

35 82. The method of producing a nucleic acid molecule as claimed in claim 81, wherein the modifying is performed by site directed mutagenesis or by use of polymerase chain reaction techniques.

83. A method of producing a recombinant vector that comprises a nucleic acid molecule that comprises a nucleotide sequence that encodes a Factor VIII:C polypeptide analog, comprising linking a regulatory 5 element to the nucleic acid molecule of claim 59.

84. A method of producing a recombinant vector that comprises a nucleic acid molecule that comprises a nucleotide sequence that encodes a Factor VIII:C 10 polypeptide analog, comprising linking a regulatory element to the nucleic acid molecule of claim 60.

85. A method of producing a recombinant vector that comprises a nucleic acid molecule that comprises a 15 nucleotide sequence that encodes a Factor VIII:C polypeptide analog, comprising linking a regulatory element to the nucleic acid molecule of claim 61.

86. A method of producing a recombinant host 20 cell that comprises a nucleic acid molecule that comprises a nucleotide sequence that encodes a Factor VIII:C polypeptide analog, comprising transforming a host cell with the nucleic acid molecule of claim 59.

25 87. A method of producing a recombinant host cell that comprises a nucleic acid molecule that comprises a nucleotide sequence that encodes a Factor VIII:C polypeptide analog, comprising transforming a host cell with the nucleic acid molecule of claim 60.

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88. A method of producing a recombinant host cell that comprises a nucleic acid molecule that comprises a nucleotide sequence that encodes a Factor VIII:C polypeptide analog, comprising transforming a host 35 cell with the nucleic acid molecule of claim 61.

89. A method of producing a recombinant host cell that comprises a recombinant vector that comprises a nucleotide sequence that encodes a Factor VIII:C polypeptide analog, comprising transforming a host cell 5 with the recombinant vector of claim 62.

90. A method of producing a recombinant host cell that comprises a recombinant vector that comprises a nucleotide sequence that encodes a Factor VIII:C 10 polypeptide analog, comprising transforming a host cell with the recombinant vector of claim 63.

91. A method of producing a recombinant host cell that comprises a recombinant vector that comprises a 15 nucleotide sequence that encodes a Factor VIII:C polypeptide analog, comprising transforming a host cell with the recombinant vector of claim 64.

92. A pharmaceutical composition comprising 20 the active Factor VIII:C polypeptide analog complex of claim 42 and a pharmaceutically acceptable excipient.

93. A pharmaceutical composition comprising the active Factor VIII:C polypeptide analog complex of 25 claim 47 and a pharmaceutically acceptable excipient.

94. A pharmaceutical composition comprising the active Factor VIII:C polypeptide analog complex of claim 52 and a pharmaceutically acceptable excipient.

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95. A method for prevention or treatment of active Factor VIII:C deficiency in a mammal comprising administering thereto a therapeutically effective amount of the pharmaceutical composition of claim 92.

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96. A method for prevention or treatment of active Factor VIII:C deficiency in a mammal comprising

administering thereto a therapeutically effective amount of the pharmaceutical composition of claim 93.

97. A method for prevention or treatment of 5 active Factor VIII:C deficiency in a mammal comprising administering thereto a therapeutically effective amount of the pharmaceutical composition of claim 94.

98. A method for prevention or treatment of 10 active Factor VIII:C deficiency in a mammal comprising administering thereto a therapeutically effective amount of the nucleic acid molecule of claim 59.

99. A method for prevention or treatment of 15 active Factor VIII:C deficiency in a mammal comprising administering thereto a therapeutically effective amount of the nucleic acid molecule of claim 60.

100. A method for prevention or treatment of 20 active Factor VIII:C deficiency in a mammal comprising administering thereto a therapeutically effective amount of the nucleic acid molecule of claim 61.

101. A method for prevention or treatment of 25 active Factor VIII:C deficiency in a mammal comprising administering thereto a therapeutically effective amount of the recombinant vector of claim 62.

102. A method for prevention or treatment of 30 active Factor VIII:C deficiency in a mammal comprising administering thereto a therapeutically effective amount of the recombinant vector of claim 63.

103. A method for prevention or treatment of 35 active Factor VIII:C deficiency in a mammal comprising administering thereto a therapeutically effective amount of the recombinant vector of claim 64.

104. A method for prevention or treatment of active Factor VIII:C deficiency in a mammal comprising administering thereto a therapeutically effective amount of the nucleic acid molecule of claim 59 and an active
5 Factor VIII:C polypeptide analog.

105. A method for prevention or treatment of active Factor VIII:C deficiency in a mammal comprising administering thereto a therapeutically effective amount
10 of the nucleic acid molecule of claim 60 and an active Factor VIII:C polypeptide analog.

106. A method for prevention or treatment of active Factor VIII:C deficiency in a mammal comprising
15 administering thereto a therapeutically effective amount of the nucleic acid molecule of claim 61 and an active Factor VIII:C polypeptide analog.

107. A method for prevention or treatment of active Factor VIII:C deficiency in a mammal comprising
20 administering thereto a therapeutically effective amount of the recombinant vector of claim 62 and an active Factor VIII:C polypeptide analog.

25 108. A method for prevention or treatment of active Factor VIII:C deficiency in a mammal comprising administering thereto a therapeutically effective amount of the recombinant vector of claim 63 and an active Factor VIII:C polypeptide analog.

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109. A method for prevention or treatment of active Factor VIII:C deficiency in a mammal comprising administering thereto a therapeutically effective amount of the recombinant vector of claim 64 and an active
35 Factor VIII:C polypeptide analog.

110. A method for prevention or treatment of active Factor VIII:C deficiency in a mammal comprising administering thereto a therapeutically effective amount of the nucleic acid molecule of claim 59 and an active
5 Factor VIII:C analog complex.

111. A method for prevention or treatment of active Factor VIII:C deficiency in a mammal comprising administering thereto a therapeutically effective amount
10 of the nucleic acid molecule of claim 60 and an active Factor VIII:C analog complex.

112. A method for prevention or treatment of active Factor VIII:C deficiency in a mammal comprising
15 administering thereto a therapeutically effective amount of the nucleic acid molecule of claim 61 and an active Factor VIII:C analog complex.

113. A method for prevention or treatment of
20 active Factor VIII:C deficiency in a mammal comprising administering thereto a therapeutically effective amount of the recombinant vector of claim 62 and an active Factor VIII:C analog complex.

25 114. A method for prevention or treatment of active Factor VIII:C deficiency in a mammal comprising administering thereto a therapeutically effective amount of the recombinant vector of claim 63 and an active Factor VIII:C analog complex.

30 35 115. A method for prevention or treatment of active Factor VIII:C deficiency in a mammal comprising administering thereto a therapeutically effective amount of the recombinant vector of claim 64 and an active Factor VIII:C analog complex.

Fig. 1A

420 Ile Gly Arg Lys Tyr Lys Val Arg Phe Met Ala Tyr Thr Asp Glu Thr Phe Lys Thr Arg Glu Ala Ile Glu Ser Gly Ile Leu Gly Pro Leu Leu Tyr Gly
 ATT GGT AGG TAC AAA AAA GTC CGA TTT ATG GCA TAC ACA GAT GAA ACC TTT AAG ACT CGT GAA CCT ATT CAG CAT GAA TCA GGA ATC TTG GGA CCT TTA CCT TAT GGG 440
 460 Glu Val Gly Asp Thr Leu Leu Ile Phe Lys Asn Gln Ala Ser Arg Pro Tyr Asn Ile Tyr Pro His Gly Ile Thr Asp Val Arg Pro Leu Tyr Ser Arg Arg Leu Pro
 GAA GTT GGA GAC ACA CTC TGG ATT ATA TTT AAG AAC CAA GCA AGC AGA CCT ATT AAC ATC TAC CCT CAC GGA ATC ACT GAT GTC CGT CCT TGT TAT TCA AGG AGA TTA CCA 480
 500 Lys Gly Val Lys His Leu Lys Asp Phe Pro Ile Leu Pro Gly Glu Ile Phe Lys Tyr Lys Icp Thr Val Thr Val Glu Asp Gly Pro Thr Lys Ser Asp Pro Arg Cys Leu
 AAA GGT GTA AAA GAT TTG AAG GAT TTT GAT CCA ATT CTG CCA ATT CTC AAA TAT AAC TGG ACA GTG ACT GTC GAA GAT GGG CCA ACT AAA TCA GAT CCT CGG TGC TGT 520
 540 Thr Arg Tyr Tyr Ser Ser Phe Val Asn Met Gly Arg Asp Leu Ala Ser Gly Leu Ile Gly Pro Leu Leu Ile Cys Tyr Lys Glu Ser Val Asp Glu Arg Gly Asn Glu Ile
 ACC CGG TAT TAC TCT AGT TTC GTC ATT ATG GAG AGA GAT CTC GCT TCA GGT TCA GGA CTC ATT GGC CCT CTC ATC TGC TAC AAA GAA TCA GAT CAA AGA GAA AAC CAG ATA 560
 570 Met Ser Asp Lys Arg Asn Val Ile Leu Phe Ser Val Phe Asp Glu Asn Arg Ser Icp Tyr Leu Thr Glu Asn Ile Glu Arg Phe Leu Pro Ala Gly Val Glu Leu
 ATG TCA GAC GAG AGG ATT GTC ATC CTG ATT GTC ATT CTA 580
 610 Glu Asp Pro Glu Phe Glu Ala Ser His Ser Ile Met His Ser Val Phe Ser Gly Tyr Val Phe Asp Ser Leu Glu Val Ser Val Cys Leu His Glu Val Ala Tyr Trp Tyr Ile Leu
 GAG GAT CCA GAG TTC CAA GCC TCC AAC ATC ATG CAC AGC ATC ATT GGC TAT ATT GAT AGT ATT GTC ATT CTA 640
 650 Ser Ile Gly Ala Glu Thr Asp Phe Leu Ser Val Phe Ser Gly Tyr Thr Phe Lys His Ile Lys Met Val Ile Gly Asp Thr Leu Phe Pro Ser Gly Glu Thr
 AGC ATT GGA GCA CAG ACT GAC TIC TIC TCT GCA TAT ACC TTC AAA AAC CAC AAA ATG GTC ATT GAA GAC AGA CTC ACC CTA TTC CCA TTC TCA GGA ACT 660
 680 Val Phe Met Ser Met Glu Asn Pro Gly Leu Trp Ile Leu Gly Cys His Asn Ser Asp Phe Arg Asn Arg Gly Met Thr Ala Leu Leu Lys Val Ser Ser Cys Asp Lys Asn
 GTC TTC ATG TCG ATG GAA AAC CCA GGT CTA TGG ATT CTG GGG TGC CAC AAC TCA GAC TTT CGG AAC AGA GGC ATG ACC GCC TTA CTC AAG GTC ATT AGT TGT GAC AAG AAC 700
 720 Thr Gly Asp Tyr Tyr Glu Asp Ser Tyr Glu Asp Ile Ser Ala Ile Tyr Leu Ser Lys Asn Asn Ala Ile Glu Pro Arg Ser Phe Ser Gln Asn Ser Arg His Pro Ser Thr
 ACT GGT GAT TAT TAC GAG GAC AGT ATT TCA GCA TAC ATT GAA GAT ATT GCA AGT AAA AAC ATT GCC ATT GAA CCT ATT GAA AGA AGC TTC TCC CAG ATT TCA AGA CCT AGC ACT 740
 760 Arg Gln Lys Glu Asn Ala Thr Thr Ile Pro Glu Asn Asp Ile Glu Lys Thr Asp Pro Trp Phe Ala His Arg Thr Pro Met Pro Lys Ile Gln Asn Val Ser Ser Ser
 AGG CAA AAG CAA TTT ATT GCC ACC ACA ATT CCA GAA ATT GAC ATA GAG ACT GAC ATT GTC ATT GCA CAC AGA ACA CCT ATT GCA AAT GTC ATT GTC ATT GTC ATT AGT 780

Fig. 1B

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Fig. 1C

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1240 Arg Gln Asn Val Glu Gly Ser Tyr Asp Gly Ala Tyr Ala Pro Val Leu Glu Asp Phe Arg Ser Leu Asn Asp Ser Thr Asn Arg Thr Lys His Thr Ala His Phe Ser
 AGG CAA AAT GTC GAA GGT TCA TAT GAC GGG GCA TAT GCT CCA GTC TTA AAT GAT TCA TTA AAT GAT TCA ACA AAT AGA ACA AAG AAG AAT GCA AAT GCT CAT TTC TCA
 1250
 1270 Lys Lys Glu Glu Glu Asn Leu Glu Gly Asn Gln Thr Lys Gln Ile Val Glu Lys Tyr Ala Cys Thr Arg Ile Ser Pro Asn Thr Ser Gln Gln Asn Phe
 GTC ACG CAA CGT AGT AAG AGA GCT TTG AAA CAA TTC AGA CTC CCA CTA GAA GAA ACA GAA CTT GAA AAA AGG ATA ATT GTG GAT GAC ACC TCA ACC CAG AGG AAT TTT
 1280
 1310 Val Thr Gln Arg Ser Lys Glu Glu Phe Arg Leu Pro Leu Glu Glu Thr Glu Leu Glu Lys Arg Ile Val ASP ASP Thr Ser Thr Gln Trp Ser Lys Asn
 Met Lys His Leu Thr Pro Ser Thr Leu Thr Gln Ile Asp Tyr Asn Glu Lys Glu Lys Gly Ala Ile Thr Gln Ser Pro Leu Ser Asp Cys Leu Thr Arg Ser His Ser Ile
 ATG AAA CAT TTG ACC CCG AGC ACC CTC ACA CAG ATA GAC TAC AAT GAG AAG GAG AAA GGG GCC ATT ACT CAG TCT CCC TTA TCA GAT TGC CTT AGG AGT CAT AGC ATC
 1320
 1350 Pro Gln Ala Asn Arg Ser Pro Ile Ala Lys Val Ser Ser Phe Pro Ser Ile Arg Pro Ile Tyr Leu Thr Arg Val Leu Phe Gln Asp Asn Ser Ser His Leu Pro
 CCT CAA GCA AAT AGG TCT CCA CCC ATT GCA AAG GTC TCA TCA TCA TCT ATT GCA TCT ATT AGA CCT ATT GCA TCT ATT GCA TCT ATT GCA TCT ATT GCA TCT ATT GCA
 1360
 1400
 1420 Ala Ala Ser Tyr Arg Lys Asp Ser Gly Val Gln Gln Ser His Phe Leu Gln Gly Ala Lys Lys Asn Asn Leu Ser Leu Ala Ile Leu Thr Leu Gln Met Thr Gly
 GCA GCA TCT ATT GCA AAG AAA GAT TCT GGG GTC CAA GAA AGC AGT CAT TTC TTA CAA GGA GCC AAA AAT AAC CTT TCT TAA GCC ATT CTA ACC TCT TGA GAG ATG ACT GGT
 1430
 1460 ASP Gln Arg Glu Val Gly Ser Leu Gln Lys Asp Leu Phe Pro Thr Glu Thr Ser Asn Gly Ser Pro Gly His Leu Asp Leu Val Gln Gln Ser Leu Gln Gln
 GAT CAA AGA GAG GTC ATT GCG TCC CTC GGG ACA AGT GTC ACA TAC AAC
 1470
 1500 Val Glu Leu Pro Lys Val His Ile Tyr Gln Lys ASP Leu Phe Pro Thr Glu Thr Ser Asn Gly Ser Pro Gly His Leu Asp Leu Val Gln Gln Ser Leu Gln Gln
 GTT GAA TTG CTT CCA AAA GTT CAC ATT TAT CAG AAG GAC CTC TCC CCT GCA ACT AGC ATT GCA
 1520
 1530 Thr Glu Gln Ala Ile Lys Trp Asn Glu Ala Asn Arg Pro Gln Lys Val Pro Phe Leu Arg Val Ala Thr Glu Ser Ser Ala Lys Thr Pro Ser Lys Leu Asp Pro Leu
 ACA GAG GCA GCG ATT AAG TGG ATT GAA GCA AAC AGA CCT GGA AAA GTT CCC TTT CTC AGA GCA ACA GAA AGC ACT CCC TCC AAG ACT CCC TCC AAG ACT CCC TCC AAG ACT CCC TCC AAG
 1540
 1570 Ala Trp Asp Asn His Tyr Gln Ile Pro Lys Glu Glu Trp Lys Ser Gln Glu Lys Ser Pro Gln Glu Lys Thr Ala Phe Lys Lys Asp Thr Ile Leu Ser Leu Asn
 GCT TGG GAT AAC CAC TAT GGT ACT CAG ATA CCA AAA GAA GAG TGG AAA TCC CAA GAG ACC TCA GCA AAA ACA GCA AAA ACA GCA AAA ACA GCA AAA ACA GCA AAA
 1580
 1610 Ala Cys Glu Ser Asn His Ala Ile Ala Ile Asn Glu Gln Asn Lys Pro Glu Ile Glu Val Thr Trp Ala Lys Gln Gly Arg Thr Glu Arg Leu Cys Ser Gln Asn
 GCT TGT GAA AGC AAT CAT GCA ATA GCA GCA ATA AAT GAG GGA CAA AAT AAG CCC GAA ATA GAA GTC ACC TGG GCA AAG CAA GTC ACC TGG GCA AAG CAA GTC ACC TGG GCA AAG CAA
 1620
 1640 Pro Pro Val Leu Lys Arg His Gln Arg Ile Thr Arg Thr Leu Gln Ser Asp Gln Glu Glu Ile Asp Tyr Asp Asp Thr Ile Ser Val Glu Met Lys Glu Asp
 CCA CCA GTC TTG AAA CGC CAT CAA CGG GAA ATA ACT CGT ACT ACR CTC ACT ACC TCA GAT TCA GAT CAA GAG GAA ATT GAC TAT GAT GAT ACC ATA TCA GAT GAA ATG AAG GAA GAA
 1650
 1660
 1670

Fig.1D

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Fig. 1E

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C12N15/12 C07K14/755 A61K38/37

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 6 C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	EP,A,0 309 237 (GENENTECH INC) 29 March 1989 see page 3, line 35 - line 51 see page 15 - page 16; example 7 ---	39
A	WO,A,87 07144 (GENETICS INST) 3 December 1987 cited in the application see page 3, paragraph 2 - page 4, paragraph 1 see page 11 - page 12 see page 17 - page 18 ---	
A	WO,A,88 08035 (GENETICS INST) 20 October 1988 see page 4, paragraph 3 - page 5, paragraph 1 -----	

Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

* Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search

28 November 1996

Date of mailing of the international search report

17.12.96

Name and mailing address of the ISA

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Authorized officer

Sitch, W

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US 96/ 11444

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. Claims Nos.: **95-115**
because they relate to subject matter not required to be searched by this Authority, namely:
Remark: Although claims 95-115 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
2. Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

The additional search fees were accompanied by the applicant's protest.

No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 96/11444

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